

STAT3

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(21) International Application Number: PCT/US00/09054 (22) International Filing Date: 6 April 2000 (06.04.00) (30) Priority Data: 09/288,461 8 April 1999 (08.04.99) US (63) Related by Continuation (CON) or Continuation-in-Part (CIP) to Earlier Application US 09/288,461 (CIP) Filed on 8 April 1999 (08.04.99) (71) Applicant (for all designated States except US): ISIS PHARMACEUTICALS, INC. [US/US]; 2292 Faraday Avenue, Carlsbad, CA 92008 (US). (72) Inventor; and (75) Inventor/Applicant (for US only): KARRAS, James, G. [US/US]; 1159 Montura Road, San Marcos, CA 92069 (US). (74) Agents: LICATA, Jane, Massey et al.; Law Offices of Jane Massey Licata, 66 E. Main Street, Marlton, NJ 08053 (US).		(81) Designated States: AE, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CR, CU, CZ, DE, DK, DM, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, TZ, UA, UG, US, UZ, VN, YU, ZA, ZW, ARIPO patent (GH, GM, KE, LS, MW, SD, SL, SZ, TZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG). Published <i>With international search report.</i>
(54) Title: ANTISENSE OLIGONUCLEOTIDE MODULATION OF STAT3 EXPRESSION (57) Abstract <p>Compounds, compositions and methods are provided for inhibiting the expression of human STAT3. The compositions comprise antisense oligonucleotides targeted to nucleic acids encoding STAT3. Methods of using these oligonucleotides for inhibition of STAT3 expression and for promotion of apoptosis are provided. Methods for treatment of diseases, particularly inflammatory diseases and cancers, associated with overexpression or constitutive activation of STAT3 or insufficient apoptosis are also provided.</p>		

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ANTISENSE OLIGONUCLEOTIDE MODULATION OF STAT3 EXPRESSION**FIELD OF THE INVENTION**

This invention relates to compositions and methods for
5 modulating expression of the human STAT3 gene, which
encodes a naturally present DNA-binding protein involved in
signal transduction and transcriptional activation, and is
implicated in disease. This invention is also directed to
methods for inhibiting STAT3-mediated signal transduction
10 and transcriptional activation; these methods can be used
diagnostically or therapeutically. Furthermore, this
invention is directed to treatment of conditions associated
with expression of the human STAT3 gene.

BACKGROUND OF THE INVENTION

15 The STAT (signal transducers and activators of
transcription) family of proteins are DNA-binding proteins
that play a dual role in signal transduction and activation
of transcription. Presently, there are six distinct
members of the STAT family (STAT1, STAT2, STAT3, STAT4,
20 STAT5, and STAT6) and several isoforms (STAT1 α , STAT1 β ,
STAT3 α and STAT3 β). The activities of the STATs are
modulated by various cytokines and mitogenic stimuli.
Binding of a cytokine to its receptor results in the
activation of Janus protein tyrosine kinases (JAKs)
25 associated with these receptors. This in turn,
phosphorylates STAT, resulting in translocation to the
nucleus and transcriptional activation of STAT responsive
genes. Phosphorylation on a specific tyrosine residue on
the STATs results in their activation, resulting in the
30 formation of homodimers and/or heterodimers of STAT which
bind to specific gene promoter sequences. Events mediated
by cytokines through STAT activation include cell
proliferation and differentiation and prevention of
apoptosis.

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The specificity of STAT activation is due to specific cytokines, i.e. each STAT is responsive to a small number of specific cytokines. Other non-cytokine signaling molecules, such as growth factors, have also been found to activate STATs. Binding of these factors to a cell surface receptor associated with protein tyrosine kinase also results in phosphorylation of STAT.

STAT3 (also acute phase response factor (APRF)), in particular, has been found to be responsive to interleukin-6 (IL-6) as well as epidermal growth factor (EGF) (Darnell, Jr., J.E., et al., *Science*, 1994, 264, 1415-1421). In addition, STAT3 has been found to have an important role in signal transduction by interferons (Yang, C.-H., et al., *Proc. Natl. Acad. Sci. USA*, 1998, 95, 5568-5572). Evidence exists suggesting that STAT3 may be regulated by the MAPK pathway. ERK2 induces serine phosphorylation and also associates with STAT3 (Jain, N., et al., *Oncogene*, 1998, 17, 3157-3167).

STAT3 is expressed in most cell types (Zhong, Z., et al., *Proc. Natl. Acad. Sci. USA*, 1994, 91, 4806-4810). It induces the expression of genes involved in response to tissue injury and inflammation. STAT3 has also been shown to prevent apoptosis through the expression of bcl-2 (Fukada, T., et al., *Immunity*, 1996, 5, 449-460).

Aberrant expression of or constitutive expression of STAT3 is associated with a number of disease processes. STAT3 has been shown to be involved in cell transformation. It is constitutively activated in v-src-transformed cells (Yu, C.-L., et al., *Science*, 1995, 269, 81-83). Constitutively active STAT3 also induces STAT3 mediated gene expression and is required for cell transformation by src (Turkson, J., et al., *Mol. Cell. Biol.*, 1998, 18, 2545-2552). STAT3 is also constitutively active in Human T cell

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lymphotropic virus I (HTLV-I) transformed cells (Migone, T.-S. et al., *Science*, 1995, 269, 79-83).

Constitutive activation and/or overexpression of STAT3 appears to be involved in several forms of cancer, including myeloma, breast carcinomas, prostate cancer, brain tumors, head and neck carcinomas, melanoma, leukemias and lymphomas. Niu et al., *Cancer Res.*, 1999, 59, 5059-5063. Breast cancer cell lines that overexpress EGFR constitutively express phosphorylated STAT3 (Sartor, C.I., et al., *Cancer Res.*, 1997, 57, 978-987; Garcia, R., et al., *Cell Growth and Differentiation*, 1997, 8, 1267-1276). Activated STAT3 levels were also found to be elevated in low grade glioblastomas and medulloblastomas (Cattaneo, E., et al., *Anticancer Res.*, 1998, 18, 2381-2387).

Cells derived from both rat and human prostate cancers have been shown to have constitutively activated STAT3, with STAT3 activation being correlated with malignant potential. Expression of a dominant-negative STAT3 was found to significantly inhibit the growth of human prostate cells. Ni et al., *Cancer Res.*, 2000, 60, 1225-1228.

STAT3 has also been found to be constitutively activated in some acute leukemias (Gouilleux-Gruart, V., et al., *Leuk. Lymphoma*, 1997, 28, 83-88) and T cell lymphoma (Yu, C.-L., et al., *J. Immunol.*, 1997, 159, 5206-5210). Interestingly, STAT3 has been found to be constitutively phosphorylated on a serine residue in chronic lymphocytic leukemia (Frank, D. A., et al., *J. Clin. Invest.*, 1997, 100, 3140-3148).

STAT3 has been found to be constitutively active in myeloma tumor cells, both in culture and in bone marrow mononuclear cells from patients with multiple myeloma. These cells are resistant to Fas-mediated apoptosis and express high levels of Bcl-xL. STAT3 signaling was shown to be essential for survival of myeloma tumor cells by

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conferring resistance to apoptosis. Thus STAT3 is a potential target for therapeutic intervention in multiple myeloma and other cancers with activated STAT3 signaling. Catlett-Falcone, R., et al., *Immunity*, 1999, 10, 105-115.

5 A gene therapy approach in a syngeneic mouse tumor model system has been used to inhibit activated STAT3 in vivo using a dominant-negative STAT3 variant. This inhibition of activated STAT3 signaling was found to suppress B16 melanoma tumor growth and induce apoptosis of B16 tumor
10 cells in vivo. Interestingly, the number of apoptotic cells (95%) exceeded the number of transfected cells, indicating a possible antitumor "bystander effect" in which an inflammatory response (tumor infiltration by acute and chronic inflammatory cells) may participate in killing of
15 residual tumor cells. Niu et al., *Cancer Res.*, 1999, 59, 5059-5063.

STAT3 may also play a role in inflammatory diseases including rheumatoid arthritis. Activated STAT3 has been found in the synovial fluid of rheumatoid arthritis
20 patients (Sengupta, T.K., et al., *J. Exp. Med.*, 1995, 181, 1015-1025) and cells from inflamed joints (Wang, F., et al., *J. Exp. Med.*, 1995, 182, 1825-1831).

Multiple forms of STAT3 exist, generated by alternative splicing. STAT3 β is a short form of STAT3
25 (also, STAT3 α) that differs predominately by the absence of 55 amino acid residues at the C-terminus. This domain contains the transactivation domain, and thus, STAT3 β may act as a negative regulator of STAT3 function (Caldenhoven, E., et al., *J. Biol. Chem.*, 1996, 271, 13221-13227).
30 STAT3 β has been found to be more stable and have greater DNA-binding activity than STAT3 α , while STAT3 α is more transcriptionally active.

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There are currently several approaches for inhibiting STAT3 expression. US Patent Nos. 5,719,042 and 5,844,082 to Akira, S. and Kishimoto, T. disclose the use of inhibitors of APRF, including antibodies, antisense nucleic acids and ribozymes for the treatment of IL-6 associated diseases, such as inflammatory diseases, leukemia, and cancer. Schreiber, R.D., et al., in US Patent Nos. 5,731,155; 5,582,999; and 5,463,023, disclose methods of inhibiting transcriptional activation using short peptides that bind p91. Darnell, J.E., et al., in US Patent No. 5,716,622, disclose peptides containing the DNA binding domain of STATs, chimeric proteins containing the DNA binding domain, and antibodies to STATs for inhibiting STAT transcriptional activation.

The use of an antisense oligonucleotide targeted to the translation start region of human STAT3 has been disclosed (Grandis, J. R., et al., *J. Clin. Invest.*, 1998, 102, 1385-1392). In this report, a phosphorothioate oligodeoxynucleotide complementary to the translation start region of STAT3 inhibited TGF- α stimulated cell growth mediated by the epidermal growth factor receptor (EGFR).

There remains an unmet need for therapeutic compositions and methods targeting expression of STAT3, and disease processes associated therewith.

SUMMARY OF THE INVENTION

The present invention provides oligonucleotides which are targeted to nucleic acids encoding STAT3 and are capable of modulating STAT3 expression. The present invention also provides chimeric oligonucleotides targeted to nucleic acids encoding human STAT3. The oligonucleotides of the invention are believed to be useful both diagnostically and therapeutically, and are believed to be particularly useful in the methods of the present invention.

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The present invention also comprises methods of modulating the expression of human STAT3, in cells and tissues, using the oligonucleotides of the invention. Methods of inhibiting STAT3 expression are provided; these
5 methods are believed to be useful both therapeutically and diagnostically. These methods are also useful as tools, for example, for detecting and determining the role of STAT3 in various cell functions and physiological processes and conditions and for diagnosing conditions associated
10 with expression of STAT3.

The present invention also comprises methods for diagnosing and treating inflammatory diseases, particularly rheumatoid arthritis, and cancers, including those of the breast, prostate, head and neck, and brain, myelomas and
15 melanomas and leukemias and lymphomas. These methods are believed to be useful, for example, in diagnosing STAT3-associated disease progression. These methods employ the oligonucleotides of the invention. These methods are believed to be useful both therapeutically, including
20 prophylactically, and as clinical research and diagnostic tools.

DETAILED DESCRIPTION OF THE INVENTION

STAT3 plays an important role in cytokine signal transduction. Overexpression and/or constitutive
25 activation of STAT3 is associated with a number of inflammatory diseases and cancers. As such, this DNA-binding protein represents an attractive target for treatment of such diseases. In particular, modulation of the expression of STAT3 may be useful for the treatment of
30 diseases such as rheumatoid arthritis, breast cancer, prostate cancer, brain cancer, head and neck cancer, myelomas, melanomas, leukemias and lymphomas.

The present invention employs antisense compounds, particularly oligonucleotides, for use in modulating the
35 function of nucleic acid molecules encoding STAT3,

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ultimately modulating the amount of STAT3 produced. This is accomplished by providing oligonucleotides which specifically hybridize with nucleic acids, preferably mRNA, encoding STAT3.

5 This relationship between an antisense compound such as an oligonucleotide and its complementary nucleic acid target, to which it hybridizes, is commonly referred to as "antisense". "Targeting" an oligonucleotide to a chosen nucleic acid target, in the context of this invention, is a
10 multistep process. The process usually begins with identifying a nucleic acid sequence whose function is to be modulated. This may be, as examples, a cellular gene (or mRNA made from the gene) whose expression is associated with a particular disease state, or a foreign nucleic acid
15 from an infectious agent. In the present invention, the targets are nucleic acids encoding STAT3; in other words, a gene encoding STAT3, or mRNA expressed from the STAT3 gene. mRNA which encodes STAT3 is presently the preferred target. The targeting process also includes determination of a site
20 or sites within the nucleic acid sequence for the antisense interaction to occur such that modulation of gene expression will result.

In accordance with this invention, persons of ordinary skill in the art will understand that messenger RNA
25 includes not only the information to encode a protein using the three letter genetic code, but also associated ribonucleotides which form a region known to such persons as the 5'-untranslated region, the 3'-untranslated region, the 5' cap region and intron/exon junction ribonucleotides.
30 Thus, oligonucleotides may be formulated in accordance with this invention which are targeted wholly or in part to these associated ribonucleotides as well as to the informational ribonucleotides. The oligonucleotide may therefore be specifically hybridizable with a transcription
35 initiation site region, a translation initiation codon

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region, a 5' cap region, an intron/exon junction, coding sequences, a translation termination codon region or sequences in the 5'- or 3'-untranslated region. Since, as is known in the art, the translation initiation codon is typically 5'-AUG (in transcribed mRNA molecules; 5'-ATG in the corresponding DNA molecule), the translation initiation codon is also referred to as the "AUG codon," the "start codon" or the "AUG start codon." A minority of genes have a translation initiation codon having the RNA sequence 5'-GUG, 5'-UUG or 5'-CUG, and 5'-AUA, 5'-ACG and 5'-CUG have been shown to function *in vivo*. Thus, the terms "translation initiation codon" and "start codon" can encompass many codon sequences, even though the initiator amino acid in each instance is typically methionine (in eukaryotes) or formylmethionine (prokaryotes). It is also known in the art that eukaryotic and prokaryotic genes may have two or more alternative start codons, any one of which may be preferentially utilized for translation initiation in a particular cell type or tissue, or under a particular set of conditions. In the context of the invention, "start codon" and "translation initiation codon" refer to the codon or codons that are used *in vivo* to initiate translation of an mRNA molecule transcribed from a gene encoding STAT3, regardless of the sequence(s) of such codons. It is also known in the art that a translation termination codon (or "stop codon") of a gene may have one of three sequences, i.e., 5'-UAA, 5'-UAG and 5'-UGA (the corresponding DNA sequences are 5'-TAA, 5'-TAG and 5'-TGA, respectively). The terms "start codon region," "AUG region" and "translation initiation codon region" refer to a portion of such an mRNA or gene that encompasses from about 25 to about 50 contiguous nucleotides in either direction (i.e., 5' or 3') from a translation initiation codon. This region is a preferred target region. Similarly, the terms "stop codon region" and "translation

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termination codon region" refer to a portion of such an mRNA or gene that encompasses from about 25 to about 50 contiguous nucleotides in either direction (i.e., 5' or 3') from a translation termination codon. This region is a preferred target region. The open reading frame (ORF) or "coding region," which is known in the art to refer to the region between the translation initiation codon and the translation termination codon, is also a region which may be targeted effectively. Other preferred target regions include the 5' untranslated region (5'UTR), known in the art to refer to the portion of an mRNA in the 5' direction from the translation initiation codon, and thus including nucleotides between the 5' cap site and the translation initiation codon of an mRNA or corresponding nucleotides on the gene and the 3' untranslated region (3'UTR), known in the art to refer to the portion of an mRNA in the 3' direction from the translation termination codon, and thus including nucleotides between the translation termination codon and 3' end of an mRNA or corresponding nucleotides on the gene. The 5' cap of an mRNA comprises an N7-methylated guanosine residue joined to the 5'-most residue of the mRNA via a 5'-5' triphosphate linkage. The 5' cap region of an mRNA is considered to include the 5' cap structure itself as well as the first 50 nucleotides adjacent to the cap. The 5' cap region may also be a preferred target region.

Although some eukaryotic mRNA transcripts are directly translated, many contain one or more regions, known as "introns", which are excised from a pre-mRNA transcript to yield one or more mature mRNA. The remaining (and therefore translated) regions are known as "exons" and are spliced together to form a continuous mRNA sequence. mRNA splice sites, i.e., exon-exon or intron-exon junctions, may also be preferred target regions, and are particularly useful in situations where aberrant splicing is implicated in disease, or where an overproduction of a particular mRNA

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splice product is implicated in disease. Aberrant fusion junctions due to rearrangements or deletions are also preferred targets. Targeting particular exons in alternatively spliced mRNAs may also be preferred. It has
5 also been found that introns can also be effective, and therefore preferred, target regions for antisense compounds targeted, for example, to DNA or pre-mRNA.

Once the target site or sites have been identified, oligonucleotides are chosen which are sufficiently
10 complementary to the target, i.e., hybridize sufficiently well and with sufficient specificity, to give the desired modulation.

"Hybridization", in the context of this invention, means hydrogen bonding, also known as Watson-Crick base
15 pairing, between complementary bases, usually on opposite nucleic acid strands or two regions of a nucleic acid strand. Guanine and cytosine are examples of complementary bases which are known to form three hydrogen bonds between them. Adenine and thymine are examples of complementary
20 bases which form two hydrogen bonds between them.

"Specifically hybridizable" and "complementary" are terms which are used to indicate a sufficient degree of complementarity such that stable and specific binding occurs between the DNA or RNA target and the
25 oligonucleotide.

It is understood that an oligonucleotide need not be 100% complementary to its target nucleic acid sequence to be specifically hybridizable. An oligonucleotide is specifically hybridizable when binding of the
30 oligonucleotide to the target interferes with the normal function of the target molecule to cause a loss of utility, and there is a sufficient degree of complementarity to avoid non-specific binding of the oligonucleotide to non-target sequences under conditions in which specific binding
35 is desired, i.e., under physiological conditions in the

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case of *in vivo* assays or therapeutic treatment or, in the case of *in vitro* assays, under conditions in which the assays are conducted.

Hybridization of antisense oligonucleotides with mRNA
5 interferes with one or more of the normal functions of mRNA. The functions of mRNA to be interfered with include all vital functions such as, for example, translocation of the RNA to the site of protein translation, translation of protein from the RNA, splicing of the RNA to yield one or
10 more mRNA species, and catalytic activity which may be engaged in by the RNA. Binding of specific protein(s) to the RNA may also be interfered with by antisense oligonucleotide hybridization to the RNA.

The overall effect of interference with mRNA function
15 is modulation of expression of STAT3. In the context of this invention "modulation" means either inhibition or stimulation; i.e., either a decrease or increase in expression. This modulation can be measured in ways which are routine in the art, for example by Northern blot assay
20 of mRNA expression, or reverse transcriptase PCR, as taught in the examples of the instant application or by Western blot or ELISA assay of protein expression, or by an immunoprecipitation assay of protein expression. Effects on cell proliferation or tumor cell growth can also be
25 measured, as taught in the examples of the instant application. Inhibition is presently preferred.

In addition to the well known antisense effects of oligonucleotides, it has also been found that oligonucleotide analogs having at least one
30 phosphorothioate bond can induce stimulation of a local immune response. This is described in U.S. Patent 5,663,153 which is commonly assigned to the assignee of the present invention and is herein incorporated by reference in its entirety. This immunostimulatory effect does not appear to
35 be related to any antisense effect which these

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oligonucleotide analogs may or may not possess. These oligonucleotide analogs are useful as immunopotentiators, either alone or in combination with other therapeutic modalities, such as drugs, particularly antiinfective and anticancer drugs, and surgical procedures to increase efficacy. In addition, the antiinfective and anticancer effects already possessed by certain antisense oligonucleotide analogs are enhanced through such immune stimulation.

10 It has also been found that oligonucleotide analogs having at least one phosphorothioate bond can be used to induce stimulation of a systemic or humoral immune response. Thus, these oligonucleotides are also useful as immunopotentiators of an antibody response, either alone or
15 in combination with other therapeutic modalities. U.S. Patent 5,663,153.

It is presently believed, therefore, that, in addition to the antisense effects of oligonucleotides targeted to STAT3, oligonucleotides containing at least one
20 phosphorothioate backbone linkage may be useful in eliciting an immune response which may add to the antitumor "bystander effect" already observed with dominant negative inhibitors of STAT3 signaling. Niu et al., *Cancer Res.*, 1999, 59, 5059-5063. This effect is believed to be related
25 to tumor infiltration by acute and chronic inflammatory cells which may participate in killing of residual tumor cells. Thus the therapeutic effects of antisense oligonucleotides targeted to STAT3 may be potentiated by the immunostimulatory properties of the oligonucleotides
30 themselves. Alternatively, oligonucleotides which may not be targeted to STAT3 but which contain at least one phosphorothioate backbone linkage may be used as adjuvants in combination with antisense or other inhibitors of STAT3.

The oligonucleotides of this invention can be used in
35 diagnostics, therapeutics, prophylaxis, and as research

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reagents and in kits. Since the oligonucleotides of this invention hybridize to nucleic acids encoding STAT3, sandwich, colorimetric and other assays can easily be constructed to exploit this fact. Provision of means for
5 detecting hybridization of oligonucleotide with the STAT3 gene or mRNA can routinely be accomplished. Such provision may include enzyme conjugation, radiolabelling or any other suitable detection systems. Kits for detecting the presence or absence of STAT3 may also be prepared.

10 The present invention is also suitable for diagnosing abnormal inflammatory states or certain cancers in tissue or other samples from patients suspected of having an inflammatory disease such as rheumatoid arthritis or cancers such as breast, brain, or head and neck cancer,
15 melanomas, myelomas, leukemias and lymphomas. A number of assays may be formulated employing the present invention, which assays will commonly comprise contacting a tissue sample with an oligonucleotide of the invention under conditions selected to permit detection and, usually,
20 quantitation of such inhibition. In the context of this invention, to "contact" tissues or cells with an oligonucleotide or oligonucleotides means to add the oligonucleotide(s), usually in a liquid carrier, to a cell suspension or tissue sample, either in vitro or ex vivo, or
25 to administer the oligonucleotide(s) to cells or tissues within an animal.

The oligonucleotides of this invention may also be used for research purposes. Thus, the specific hybridization exhibited by the oligonucleotides may be used
30 for assays, purifications, cellular product preparations and in other methodologies which may be appreciated by persons of ordinary skill in the art.

In the context of this invention, the term "oligonucleotide" refers to an oligomer or polymer of
35 ribonucleic acid or deoxyribonucleic acid. This term

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includes oligonucleotides composed of naturally-occurring nucleobases, sugars and covalent intersugar (backbone) linkages as well as oligonucleotides having non-naturally-occurring portions which function similarly. Such modified or substituted oligonucleotides are often preferred over native forms because of desirable properties such as, for example, enhanced cellular uptake, enhanced binding to target and increased stability in the presence of nucleases.

10 The antisense compounds in accordance with this invention preferably comprise from about 5 to about 50 nucleobases. Particularly preferred are antisense oligonucleotides comprising from about 8 to about 30 nucleobases (i.e. from about 8 to about 30 linked
15 nucleosides). As is known in the art, a nucleoside is a base-sugar combination. The base portion of the nucleoside is normally a heterocyclic base. The two most common classes of such heterocyclic bases are the purines and the pyrimidines. Nucleotides are nucleosides that further
20 include a phosphate group covalently linked to the sugar portion of the nucleoside. For those nucleosides that include a pentofuranosyl sugar, the phosphate group can be linked to either the 2', 3' or 5' hydroxyl moiety of the sugar. In forming oligonucleotides, the phosphate groups
25 covalently link adjacent nucleosides to one another to form a linear polymeric compound. In turn the respective ends of this linear polymeric structure can be further joined to form a circular structure, however, open linear structures are generally preferred. Within the oligonucleotide
30 structure, the phosphate groups are commonly referred to as forming the internucleoside backbone of the oligonucleotide. The normal linkage or backbone of RNA and DNA is a 3' to 5' phosphodiester linkage.

Specific examples of preferred antisense compounds
35 useful in this invention include oligonucleotides

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containing modified backbones or non-natural internucleoside linkages. As defined in this specification, oligonucleotides having modified backbones include those that retain a phosphorus atom in the backbone and those that do not have a phosphorus atom in the backbone. For the purposes of this specification, and as sometimes referenced in the art, modified oligonucleotides that do not have a phosphorus atom in their internucleoside backbone can also be considered to be oligonucleosides.

Preferred modified oligonucleotide backbones include, for example, phosphorothioates, chiral phosphorothioates, phosphorodithioates, phosphotriesters, aminoalkylphosphotriesters, methyl and other alkyl phosphonates including 3'-alkylene phosphonates and chiral phosphonates, phosphinates, phosphoramidates including 3'-amino phosphoramidate and aminoalkylphosphoramidates, thionophosphoramidates, thionoalkylphosphonates, thionoalkylphosphotriesters, and boranophosphates having normal 3'-5' linkages, 2'-5' linked analogs of these, and those having inverted polarity wherein the adjacent pairs of nucleoside units are linked 3'-5' to 5'-3' or 2'-5' to 5'-2'. Various salts, mixed salts and free acid forms are also included.

Representative United States patents that teach the preparation of the above phosphorus-containing linkages include, but are not limited to, U.S.: 3,687,808; 4,469,863; 4,476,301; 5,023,243; 5,177,196; 5,188,897; 5,264,423; 5,276,019; 5,278,302; 5,286,717; 5,321,131; 5,399,676; 5,405,939; 5,453,496; 5,455,233; 5,466,677; 5,476,925; 5,519,126; 5,536,821; 5,541,306; 5,550,111; 5,563,253; 5,571,799; 5,587,361; and 5,625,050.

Preferred modified oligonucleotide backbones that do not include a phosphorus atom therein have backbones that are formed by short chain alkyl or cycloalkyl internucleoside linkages, mixed heteroatom and alkyl or

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cycloalkyl internucleoside linkages, or one or more short chain heteroatomic or heterocyclic internucleoside linkages. These include those having morpholino linkages (formed in part from the sugar portion of a nucleoside);

5 siloxane backbones; sulfide, sulfoxide and sulfone backbones; formacetyl and thioformacetyl backbones; methylene formacetyl and thioformacetyl backbones; alkene containing backbones; sulfamate backbones; methyleneimino and methylenehydrazino backbones; sulfonate and sulfonamide

10 backbones; amide backbones; and others having mixed N, O, S and CH₂ component parts.

Representative United States patents that teach the preparation of the above oligonucleosides include, but are not limited to, U.S.: 5,034,506; 5,166,315; 5,185,444;

15 5,214,134; 5,216,141; 5,235,033; 5,264,562; 5,264,564; 5,405,938; 5,434,257; 5,466,677; 5,470,967; 5,489,677; 5,541,307; 5,561,225; 5,596,086; 5,602,240; 5,610,289; 5,602,240; 5,608,046; 5,610,289; 5,618,704; 5,623,070; 5,663,312; 5,633,360; 5,677,437; and 5,677,439.

20 In other preferred oligonucleotide mimetics, both the sugar and the internucleoside linkage, i.e., the backbone, of the nucleotide units are replaced with novel groups. The base units are maintained for hybridization with an appropriate nucleic acid target compound. One such

25 oligomeric compound, an oligonucleotide mimetic that has been shown to have excellent hybridization properties, is referred to as a peptide nucleic acid (PNA). In PNA compounds, the sugar-backbone of an oligonucleotide is replaced with an amide containing backbone, in particular

30 an aminoethylglycine backbone. The nucleobases are retained and are bound directly or indirectly to nitrogen atoms of the amide portion of the backbone. Representative United States patents that teach the preparation of PNA compounds include, but are not limited

35 to, U.S.: 5,539,082; 5,714,331; and 5,719,262. Further

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teaching of PNA compounds can be found in Nielsen et al. (Science, 1991, 254, 1497-1500).

Most preferred embodiments of the invention are oligonucleotides with phosphorothioate backbones and
5 oligonucleosides with heteroatom backbones, and in particular $-\text{CH}_2-\text{NH}-\text{O}-\text{CH}_2-$, $-\text{CH}_2-\text{N}(\text{CH}_3)-\text{O}-\text{CH}_2-$ [known as a methylene (methylimino) or MMI backbone], $-\text{CH}_2-\text{O}-\text{N}(\text{CH}_3)-\text{CH}_2-$, $-\text{CH}_2-\text{N}(\text{CH}_3)-\text{N}(\text{CH}_3)-\text{CH}_2-$ and $-\text{O}-\text{N}(\text{CH}_3)-\text{CH}_2-\text{CH}_2-$ [wherein the native phosphodiester backbone is represented as $-\text{O}-\text{P}-\text{O}-$
10 CH_2-] of the above referenced U.S. patent 5,489,677, and the amide backbones of the above referenced U.S. patent 5,602,240. Also preferred are oligonucleotides having morpholino backbone structures of the above-referenced U.S. patent 5,034,506.

15 Modified oligonucleotides may also contain one or more substituted sugar moieties. Preferred oligonucleotides comprise one of the following at the 2' position: OH; F; O-, S-, or N-alkyl, O-alkyl-O-alkyl, O-, S-, or N-alkenyl, or O-, S- or N-alkynyl, wherein the alkyl, alkenyl and alkynyl
20 may be substituted or unsubstituted C_1 to C_{10} alkyl or C_2 to C_{10} alkenyl and alkynyl. Particularly preferred are $\text{O}[(\text{CH}_2)_n\text{O}]_m\text{CH}_3$, $\text{O}(\text{CH}_2)_n\text{OCH}_3$, $\text{O}(\text{CH}_2)_2\text{ON}(\text{CH}_3)_2$, $\text{O}(\text{CH}_2)_n\text{NH}_2$, $\text{O}(\text{CH}_2)_n\text{CH}_3$, $\text{O}(\text{CH}_2)_n\text{ONH}_2$, and $\text{O}(\text{CH}_2)_n\text{ON}[(\text{CH}_2)_n\text{CH}_3]_2$, where n and m are from 1 to about 10. Other preferred oligonucleotides
25 comprise one of the following at the 2' position: C_1 to C_{10} lower alkyl, substituted lower alkyl, alkaryl, aralkyl, O-alkaryl or O-aralkyl, SH, SCH_3 , OCN, Cl, Br, CN, CF_3 , OCF_3 , SOCH_3 , SO_2CH_3 , ONO_2 , NO_2 , N_3 , NH_2 , heterocycloalkyl, heterocycloalkaryl, aminoalkylamino, polyalkylamino,
30 substituted silyl, an RNA cleaving group, a reporter group, an intercalator, a group for improving the pharmacokinetic properties of an oligonucleotide, or a group for improving the pharmacodynamic properties of an oligonucleotide, and other substituents having similar properties. A preferred
35 modification includes 2'-methoxyethoxy ($2'-\text{O}-\text{CH}_2\text{CH}_2\text{OCH}_3$, also

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known as 2'-O-(2-methoxyethyl) or 2'-MOE) (Martin et al., *Helv. Chim. Acta* 1995, 78, 486-504) i.e., an alkoxyalkoxy group. A further preferred modification includes 2'-dimethylaminooxyethoxy, i.e., a $O(CH_2)_2ON(CH_3)_2$ group, also known as 2'-DMAOE, as described in examples hereinbelow, and 2'-dimethylaminoethoxyethoxy (also known in the art as 2'-O-dimethylaminoethoxyethyl or 2'-DMAEOE), i.e., $2'-O-CH_2-O-CH_2-N(CH_2)_2$, also described in examples hereinbelow.

Other preferred modifications include 2'-methoxy (2'-O-CH₃), 2'-aminopropoxy (2'-OCH₂CH₂CH₂NH₂) and 2'-fluoro (2'-F). Similar modifications may also be made at other positions on the oligonucleotide, particularly the 3' position of the sugar on the 3' terminal nucleotide or in 2'-5' linked oligonucleotides and the 5' position of 5' terminal nucleotide. Oligonucleotides may also have sugar mimetics such as cyclobutyl moieties in place of the pentofuranosyl sugar. Representative United States patents that teach the preparation of such modified sugars structures include, but are not limited to, U.S.:

4,981,957; 5,118,800; 5,319,080; 5,359,044; 5,393,878;
5,446,137; 5,466,786; 5,514,785; 5,519,134; 5,567,811;
5,576,427; 5,591,722; 5,597,909; 5,610,300; 5,627,0531
5,639,873; 5,646,265; 5,658,873; 5,670,633; and 5,700,920.

Oligonucleotides may also include nucleobase (often referred to in the art simply as "base") modifications or substitutions. As used herein, "unmodified" or "natural" nucleobases include the purine bases adenine (A) and guanine (G), and the pyrimidine bases thymine (T), cytosine (C) and uracil (U). Modified nucleobases include other synthetic and natural nucleobases such as 5-methylcytosine (5-me-C or m5c), 5-hydroxymethyl cytosine, xanthine, hypoxanthine, 2-aminoadenine, 6-methyl and other alkyl derivatives of adenine and guanine, 2-propyl and other alkyl derivatives of adenine and guanine, 2-thiouracil, 2-

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thiothymine and 2-thiocytosine, 5-halouracil and cytosine, 5-propynyl uracil and cytosine, 6-azo uracil, cytosine and thymine, 5-uracil (pseudouracil), 4-thiouracil, 8-halo, 8-amino, 8-thiol, 8-thioalkyl, 8-hydroxyl and other 8-substituted adenines and guanines, 5-halo particularly 5-bromo, 5-trifluoromethyl and other 5-substituted uracils and cytosines, 7-methylguanine and 7-methyladenine, 8-azaguanine and 8-azaadenine, 7-deazaguanine and 7-deazaadenine and 3-deazaguanine and 3-deazaadenine.

10 Further nucleobases include those disclosed in United States Patent No. 3,687,808, those disclosed in the *Concise Encyclopedia Of Polymer Science And Engineering 1990*, pages 858-859, Kroschwitz, J.I., ed. John Wiley & Sons, those disclosed by Englisch et al. (*Angewandte Chemie, International Edition 1991*, 30, 613-722), and those

15 disclosed by Sanghvi, Y.S., Chapter 15, *Antisense Research and Applications 1993*, pages 289-302, Crooke, S.T. and Lebleu, B., ed., CRC Press. Certain of these nucleobases are particularly useful for increasing the binding affinity

20 of the oligomeric compounds of the invention. These include 5-substituted pyrimidines, 6-azapyrimidines and N-2, N-6 and O-6 substituted purines, including 2-aminopropyladenine, 5-propynyluracil and 5-propynylcytosine. 5-methylcytosine substitutions have been

25 shown to increase nucleic acid duplex stability by 0.6-1.2°C (Sanghvi, Y.S., Crooke, S.T. and Lebleu, B., eds., *Antisense Research and Applications 1993*, CRC Press, Boca Raton, pages 276-278) and are presently preferred base substitutions, even more particularly when combined with

30 2'-O-methoxyethyl sugar modifications.

Representative United States patents that teach the preparation of certain of the above noted modified nucleobases as well as other modified nucleobases include, but are not limited to, the above noted U.S. 3,687,808, as

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well as U.S.: 4,845,205; 5,130,302; 5,134,066; 5,175,273; 5,367,066; 5,432,272; 5,457,187; 5,459,255; 5,484,908; 5,502,177; 5,525,711; 5,552,540; 5,587,469; 5,594,121, 5,596,091; 5,614,617; and 5,681,941.

5 Another modification of the oligonucleotides of the invention involves chemically linking to the oligonucleotide one or more moieties or conjugates which enhance the activity, cellular distribution or cellular uptake of the oligonucleotide. Such moieties include but
10 are not limited to lipid moieties such as a cholesterol moiety (Letsinger et al., *Proc. Natl. Acad. Sci. USA* 1989, 86, 6553-6556), cholic acid (Manoharan et al., *Bioorg. Med. Chem. Lett.* 1994, 4, 1053-1059), a thioether, e.g., hexyl-S-tritylthiol (Manoharan et al., *Ann. N.Y. Acad. Sci.* 1992, 15 660, 306-309; Manoharan et al., *Bioorg. Med. Chem. Lett.* 1993, 3, 2765-2770), a thiocholesterol (Oberhauser et al., *Nucl. Acids Res.* 1992, 20, 533-538), an aliphatic chain, e.g., dodecandiol or undecyl residues (Saison-Behmoaras et al., *EMBO J.* 1991, 10, 1111-1118; Kabanov et al., *FEBS*
20 *Lett.* 1990, 259, 327-330; Svinarchuk et al., *Biochimie* 1993, 75, 49-54), a phospholipid, e.g., di-hexadecyl-rac-glycerol or triethylammonium 1,2-di-O-hexadecyl-rac-glycero-3-H-phosphonate (Manoharan et al., *Tetrahedron Lett.* 1995, 36, 3651-3654; Shea et al., *Nucl. Acids Res.*
25 1990, 18, 3777-3783), a polyamine or a polyethylene glycol chain (Manoharan et al., *Nucleosides & Nucleotides* 1995, 14, 969-973), or adamantane acetic acid (Manoharan et al., *Tetrahedron Lett.* 1995, 36, 3651-3654), a palmityl moiety (Mishra et al., *Biochim. Biophys. Acta* 1995, 1264, 229-
30 237), or an octadecylamine or hexylamino-carbonyl-oxycholesterol moiety (Crooke et al., *J. Pharmacol. Exp. Ther.* 1996, 277, 923-937).

Representative United States patents that teach the preparation of such oligonucleotide conjugates include, but

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are not limited to, U.S.: 4,828,979; 4,948,882; 5,218,105;
5,525,465; 5,541,313; 5,545,730; 5,552,538; 5,578,717,
5,580,731; 5,580,731; 5,591,584; 5,109,124; 5,118,802;
5,138,045; 5,414,077; 5,486,603; 5,512,439; 5,578,718;
5 5,608,046; 4,587,044; 4,605,735; 4,667,025; 4,762,779;
4,789,737; 4,824,941; 4,835,263; 4,876,335; 4,904,582;
4,958,013; 5,082,830; 5,112,963; 5,214,136; 5,082,830;
5,112,963; 5,214,136; 5,245,022; 5,254,469; 5,258,506;
5,262,536; 5,272,250; 5,292,873; 5,317,098; 5,371,241,
10 5,391,723; 5,416,203; 5,451,463; 5,510,475; 5,512,667;
5,514,785; 5,565,552; 5,567,810; 5,574,142; 5,585,481;
5,587,371; 5,595,726; 5,597,696; 5,599,923; 5,599,928 and
5,688,941.

The present invention also includes oligonucleotides
15 which are chimeric oligonucleotides. "Chimeric"
oligonucleotides or "chimeras," in the context of this
invention, are oligonucleotides which contain two or more
chemically distinct regions, each made up of at least one
nucleotide. These oligonucleotides typically contain at
20 least one region wherein the oligonucleotide is modified so
as to confer upon the oligonucleotide increased resistance
to nuclease degradation, increased cellular uptake, and/or
increased binding affinity for the target nucleic acid. An
additional region of the oligonucleotide may serve as a
25 substrate for enzymes capable of cleaving RNA:DNA or
RNA:RNA hybrids. By way of example, RNase H is a cellular
endonuclease which cleaves the RNA strand of an RNA:DNA
duplex. Activation of RNase H, therefore, results in
cleavage of the RNA target, thereby greatly enhancing the
30 efficiency of antisense inhibition of gene expression.
Cleavage of the RNA target can be routinely detected by gel
electrophoresis and, if necessary, associated nucleic acid
hybridization techniques known in the art. This RNase H-
mediated cleavage of the RNA target is distinct from the

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use of ribozymes to cleave nucleic acids. Ribozymes are not comprehended by the present invention.

Examples of chimeric oligonucleotides include but are not limited to "gapmers," in which three distinct regions are present, normally with a central region flanked by two regions which are chemically equivalent to each other but distinct from the gap. A preferred example of a gapmer is an oligonucleotide in which a central portion (the "gap") of the oligonucleotide serves as a substrate for RNase H and is preferably composed of 2'-deoxynucleotides, while the flanking portions (the 5' and 3' "wings") are modified to have greater affinity for the target RNA molecule but are unable to support nuclease activity (e.g., fluoro- or 2'-O-methoxyethyl- substituted). Chimeric oligonucleotides are not limited to those with modifications on the sugar, but may also include oligonucleosides or oligonucleotides with modified backbones, e.g., with regions of phosphorothioate (P=S) and phosphodiester (P=O) backbone linkages or with regions of MMI and P=S backbone linkages. Other chimeras include "wingmers," also known in the art as "hemimers," that is, oligonucleotides with two distinct regions. In a preferred example of a wingmer, the 5' portion of the oligonucleotide serves as a substrate for RNase H and is preferably composed of 2'-deoxynucleotides, whereas the 3' portion is modified in such a fashion so as to have greater affinity for the target RNA molecule but is unable to support nuclease activity (e.g., 2'-fluoro- or 2'-O-methoxyethyl- substituted), or vice-versa. In one embodiment, the oligonucleotides of the present invention contain a 2'-O-methoxyethyl (2'-O-CH₂CH₂OCH₃) modification on the sugar moiety of at least one nucleotide. This modification has been shown to increase both affinity of the oligonucleotide for its target and nuclease resistance of the oligonucleotide. According to the invention, one, a plurality, or all of the nucleotide subunits of the

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oligonucleotides of the invention may bear a 2'-O-methoxyethyl (-O-CH₂CH₂OCH₃) modification. Oligonucleotides comprising a plurality of nucleotide subunits having a 2'-O-methoxyethyl modification can have such a modification on
5 any of the nucleotide subunits within the oligonucleotide, and may be chimeric oligonucleotides. Aside from or in addition to 2'-O-methoxyethyl modifications, oligonucleotides containing other modifications which enhance antisense efficacy, potency or target affinity are
10 also preferred. Chimeric oligonucleotides comprising one or more such modifications are presently preferred.

The oligonucleotides used in accordance with this invention may be conveniently and routinely made through the well-known technique of solid phase synthesis.
15 Equipment for such synthesis is sold by several vendors including Applied Biosystems. Any other means for such synthesis may also be employed; the actual synthesis of the oligonucleotides is well within the talents of the routineer. It is well known to use similar techniques to
20 prepare oligonucleotides such as the phosphorothioates and 2'-alkoxy or 2'-alkoxyalkoxy derivatives, including 2'-O-methoxyethyl oligonucleotides (Martin, P., *Helv. Chim. Acta* 1995, 78, 486-504). It is also well known to use similar techniques and commercially available modified amidites and
25 controlled-pore glass (CPG) products such as biotin, fluorescein, acridine or psoralen-modified amidites and/or CPG (available from Glen Research, Sterling, VA) to synthesize fluorescently labeled, biotinylated or other conjugated oligonucleotides.

30 The antisense compounds of the present invention include bioequivalent compounds, including pharmaceutically acceptable salts and prodrugs. This is intended to encompass any pharmaceutically acceptable salts, esters, or salts of such esters, or any other compound which, upon
35 administration to an animal including a human, is capable

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of providing (directly or indirectly) the biologically active metabolite or residue thereof. Accordingly, for example, the disclosure is also drawn to pharmaceutically acceptable salts of the nucleic acids of the invention and
5 prodrugs of such nucleic acids. "Pharmaceutically acceptable salts" are physiologically and pharmaceutically acceptable salts of the nucleic acids of the invention: i.e., salts that retain the desired biological activity of the parent compound and do not impart undesired
10 toxicological effects thereto (see, for example, Berge et al., "Pharmaceutical Salts," *J. of Pharma Sci.* 1977, 66, 1-19).

For oligonucleotides, examples of pharmaceutically acceptable salts include but are not limited to (a) salts
15 formed with cations such as sodium, potassium, ammonium, magnesium, calcium, polyamines such as spermine and spermidine, etc.; (b) acid addition salts formed with inorganic acids, for example hydrochloric acid, hydrobromic acid, sulfuric acid, phosphoric acid, nitric acid and the
20 like; (c) salts formed with organic acids such as, for example, acetic acid, oxalic acid, tartaric acid, succinic acid, maleic acid, fumaric acid, gluconic acid, citric acid, malic acid, ascorbic acid, benzoic acid, tannic acid, palmitic acid, alginic acid, polyglutamic acid,
25 naphthalenesulfonic acid, methanesulfonic acid, p-toluenesulfonic acid, naphthalenedisulfonic acid, polygalacturonic acid, and the like; and (d) salts formed from elemental anions such as chlorine, bromine, and iodine.

30 The oligonucleotides of the invention may additionally or alternatively be prepared to be delivered in a "prodrug" form. The term "prodrug" indicates a therapeutic agent that is prepared in an inactive form that is converted to an active form (i.e., drug) within the body or cells thereof
35 by the action of endogenous enzymes or other chemicals

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and/or conditions. In particular, prodrug versions of the oligonucleotides of the invention are prepared as SATE [(S-acetyl-2-thioethyl) phosphate] derivatives according to the methods disclosed in WO 93/24510 to Gosselin et al.,
5 published December 9, 1993.

For therapeutic or prophylactic treatment, oligonucleotides are administered in accordance with this invention. Oligonucleotide compounds of the invention may be formulated in a pharmaceutical composition, which may
10 include pharmaceutically acceptable carriers, thickeners, diluents, buffers, preservatives, surface active agents, neutral or cationic lipids, lipid complexes, liposomes, penetration enhancers, carrier compounds and other pharmaceutically acceptable carriers or excipients and the
15 like in addition to the oligonucleotide. Such compositions and formulations are comprehended by the present invention.

Pharmaceutical compositions comprising the oligonucleotides of the present invention may include penetration enhancers in order to enhance the alimentary
20 delivery of the oligonucleotides. Penetration enhancers may be classified as belonging to one of five broad categories, i.e., fatty acids, bile salts, chelating agents, surfactants and non-surfactants (Lee et al., *Critical Reviews in Therapeutic Drug Carrier Systems* 1991, 8,
25 91-192; Muranishi, *Critical Reviews in Therapeutic Drug Carrier Systems* 1990, 7, 1-33). One or more penetration enhancers from one or more of these broad categories may be included. Various fatty acids and their derivatives which act as penetration enhancers include, for example,
30 oleic acid, lauric acid, capric acid, myristic acid, palmitic acid, stearic acid, linoleic acid, linolenic acid, dicaprate, tricaprate, recinleate, monoolein (a.k.a. 1-monooleoyl-rac-glycerol), dilaurin, caprylic acid, arachidonic acid, glyceryl 1-monocaprate,
35 1-dodecylazacycloheptan-2-one, acylcarnitines,

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acylcholines, mono- and di-glycerides and physiologically acceptable salts thereof (i.e., oleate, laurate, caprate, myristate, palmitate, stearate, linoleate, etc.) (Lee et al., *Critical Reviews in Therapeutic Drug Carrier Systems* 5 1991, page 92; Muranishi, *Critical Reviews in Therapeutic Drug Carrier Systems* 1990, 7, 1; El-Hariri et al., *J. Pharm. Pharmacol.* 1992 44, 651-654).

The physiological roles of bile include the facilitation of dispersion and absorption of lipids and fat-soluble vitamins (Brunton, Chapter 38 In: Goodman & Gilman's *The Pharmacological Basis of Therapeutics*, 9th Ed., Hardman et al., eds., McGraw-Hill, New York, NY, 1996, pages 934-935). Various natural bile salts, and their synthetic derivatives, act as penetration enhancers. Thus, 15 the term "bile salt" includes any of the naturally occurring components of bile as well as any of their synthetic derivatives.

Complex formulations comprising one or more penetration enhancers may be used. For example, bile salts 20 may be used in combination with fatty acids to make complex formulations.

Chelating agents include, but are not limited to, disodium ethylenediaminetetraacetate (EDTA), citric acid, salicylates (e.g., sodium salicylate, 5-methoxysalicylate 25 and homovanilate), N-acyl derivatives of collagen, laureth-9 and N-amino acyl derivatives of beta-diketones (enamines) [Lee et al., *Critical Reviews in Therapeutic Drug Carrier Systems* 1991, page 92; Muranishi, *Critical Reviews in Therapeutic Drug Carrier Systems* 1990, 7, 1-33; Buur et al., *J. Control Rel.* 1990, 14, 43-51). Chelating agents 30 have the added advantage of also serving as DNase inhibitors.

Surfactants include, for example, sodium lauryl sulfate, polyoxyethylene-9-lauryl ether and

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polyoxyethylene-20-cetyl ether (Lee et al., *Critical Reviews in Therapeutic Drug Carrier Systems* 1991, page 92); and perfluorochemical emulsions, such as FC-43 (Takahashi et al., *J. Pharm. Pharmacol.* 1988, 40, 252-257).

- 5 Non-surfactants include, for example, unsaturated cyclic ureas, 1-alkyl- and 1-alkenylazacyclo-alkanone derivatives (Lee et al., *Critical Reviews in Therapeutic Drug Carrier Systems* 1991, page 92); and non-steroidal anti-inflammatory agents such as diclofenac sodium,
10 indomethacin and phenylbutazone (Yamashita et al., *J. Pharm. Pharmacol.* 1987, 39, 621-626).

As used herein, "carrier compound" refers to a nucleic acid, or analog thereof, which is inert (i.e., does not possess biological activity *per se*) but is recognized as a
15 nucleic acid by *in vivo* processes that reduce the bioavailability of a nucleic acid having biological activity by, for example, degrading the biologically active nucleic acid or promoting its removal from circulation. The coadministration of a nucleic acid and a carrier
20 compound, typically with an excess of the latter substance, can result in a substantial reduction of the amount of nucleic acid recovered in the liver, kidney or other extracirculatory reservoirs, presumably due to competition between the carrier compound and the nucleic acid for a
25 common receptor. In contrast to a carrier compound, a "pharmaceutically acceptable carrier" (excipient) is a pharmaceutically acceptable solvent, suspending agent or any other pharmacologically inert vehicle for delivering one or more nucleic acids to an animal. The
30 pharmaceutically acceptable carrier may be liquid or solid and is selected with the planned manner of administration in mind so as to provide for the desired bulk, consistency, etc., when combined with a nucleic acid and the other components of a given pharmaceutical composition. Typical

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pharmaceutically acceptable carriers include, but are not limited to, binding agents (e.g., pregelatinized maize starch, polyvinylpyrrolidone or hydroxypropyl methylcellulose, etc.); fillers (e.g., lactose and other
5 sugars, microcrystalline cellulose, pectin, gelatin, calcium sulfate, ethyl cellulose, polyacrylates or calcium hydrogen phosphate, etc.); lubricants (e.g., magnesium stearate, talc, silica, colloidal silicon dioxide, stearic acid, metallic stearates, hydrogenated vegetable oils, corn
10 starch, polyethylene glycols, sodium benzoate, sodium acetate, etc.); disintegrates (e.g., starch, sodium starch glycolate, etc.); or wetting agents (e.g., sodium lauryl sulphate, etc.). Sustained release oral delivery systems and/or enteric coatings for orally administered dosage
15 forms are described in U.S. Patents Nos. 4,704,295; 4,556,552; 4,309,406; and 4,309,404.

The compositions of the present invention may additionally contain other adjunct components conventionally found in pharmaceutical compositions, at
20 their art-established usage levels. Thus, for example, the compositions may contain additional compatible pharmaceutically-active materials such as, e.g., antipruritics, astringents, local anesthetics or anti-inflammatory agents, or may contain additional
25 materials useful in physically formulating various dosage forms of the composition of present invention, such as dyes, flavoring agents, preservatives, antioxidants, opacifiers, thickening agents and stabilizers. However, such materials, when added, should not unduly interfere
30 with the biological activities of the components of the compositions of the invention.

Regardless of the method by which the oligonucleotides of the invention are introduced into a patient, colloidal dispersion systems may be used as delivery vehicles to
35 enhance the *in vivo* stability of the oligonucleotides

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and/or to target the oligonucleotides to a particular organ, tissue or cell type. Colloidal dispersion systems include, but are not limited to, macromolecule complexes, nanocapsules, microspheres, beads and lipid-based systems including oil-in-water emulsions, micelles, mixed micelles, liposomes and lipid:oligonucleotide complexes of uncharacterized structure. A preferred colloidal dispersion system is a plurality of liposomes. Liposomes are microscopic spheres having an aqueous core surrounded by one or more outer layers made up of lipids arranged in a bilayer configuration (see, generally, Chonn et al., *Current Op. Biotech.* 1995, 6, 698-708).

The pharmaceutical compositions of the present invention may be administered in a number of ways depending upon whether local or systemic treatment is desired and upon the area to be treated. Administration may be topical (including ophthalmic, vaginal, rectal, intranasal, epidermal, and transdermal), oral or parenteral. Parenteral administration includes intravenous drip, subcutaneous, intraperitoneal or intramuscular injection, pulmonary administration, e.g., by inhalation or insufflation, or intracranial, e.g., intrathecal or intraventricular, administration. Oligonucleotides with at least one 2'-O-methoxyethyl modification are believed to be particularly useful for oral administration.

Formulations for topical administration may include transdermal patches, ointments, lotions, creams, gels, drops, suppositories, sprays, liquids and powders. Conventional pharmaceutical carriers, aqueous, powder or oily bases, thickeners and the like may be necessary or desirable. Coated condoms, gloves and the like may also be useful.

Compositions for oral administration include powders or granules, suspensions or solutions in water or non-aqueous media, capsules, sachets or tablets. Thickeners,

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flavoring agents, diluents, emulsifiers, dispersing aids or binders may be desirable.

Compositions for parenteral administration may include sterile aqueous solutions which may also contain buffers, diluents and other suitable additives. In some cases it may be more effective to treat a patient with an oligonucleotide of the invention in conjunction with other traditional therapeutic modalities in order to increase the efficacy of a treatment regimen. In the context of the invention, the term "treatment regimen" is meant to encompass therapeutic, palliative and prophylactic modalities. For example, a patient may be treated with conventional chemotherapeutic agents, particularly those used for tumor and cancer treatment. Examples of such chemotherapeutic agents include but are not limited to daunorubicin, daunomycin, dactinomycin, doxorubicin, epirubicin, idarubicin, esorubicin, bleomycin, mafosfamide, ifosfamide, cytosine arabinoside, bis-chloroethylnitrosurea, busulfan, mitomycin C, actinomycin D, mithramycin, prednisone, hydroxyprogesterone, testosterone, tamoxifen, dacarbazine, procarbazine, hexamethylmelamine, pentamethylmelamine, mitoxantrone, amsacrine, chlorambucil, methylcyclohexylnitrosurea, nitrogen mustards, melphalan, cyclophosphamide, 6-mercaptapurine, 6-thioguanine, cytarabine (CA), 5-azacytidine, hydroxyurea, deoxycoformycin, 4-hydroxyperoxycyclophosphoramidate, 5-fluorouracil (5-FU), 5-fluorodeoxyuridine (5-FUdR), methotrexate (MTX), colchicine, taxol, vincristine, vinblastine, etoposide, trimetrexate, teniposide, cisplatin and diethylstilbestrol (DES). See, generally, *The Merck Manual of Diagnosis and Therapy*, 15th Ed. 1987, pp. 1206-1228, Berkow et al., eds., Rahway, N.J. When used with the compounds of the invention, such chemotherapeutic agents may be used individually (e.g., 5-FU and oligonucleotide), sequentially

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(e.g., 5-FU and oligonucleotide for a period of time followed by MTX and oligonucleotide), or in combination with one or more other such chemotherapeutic agents (e.g., 5-FU, MTX and oligonucleotide, or 5-FU, radiotherapy and
5 oligonucleotide).

The formulation of therapeutic compositions and their subsequent administration is believed to be within the skill of those in the art. Dosing is dependent on severity and responsiveness of the disease state to be treated, with
10 the course of treatment lasting from several days to several months, or until a cure is effected or a diminution of the disease state is achieved. Optimal dosing schedules can be calculated from measurements of drug accumulation in the body of the patient. Persons of ordinary skill can
15 easily determine optimum dosages, dosing methodologies and repetition rates. Optimum dosages may vary depending on the relative potency of individual oligonucleotides, and can generally be estimated based on EC_{50} s found to be effective *in vitro* and in *in vivo* animal models. In
20 general, dosage is from 0.01 μ g to 100 g per kg of body weight, and may be given once or more daily, weekly, monthly or yearly, or even once every 2 to 20 years. Persons of ordinary skill in the art can easily estimate repetition rates for dosing based on measured residence
25 times and concentrations of the drug in bodily fluids or tissues. Following successful treatment, it may be desirable to have the patient undergo maintenance therapy to prevent the recurrence of the disease state, wherein the oligonucleotide is administered in maintenance doses,
30 ranging from 0.01 μ g to 100 g per kg of body weight, once or more daily, to once every 20 years.

The following examples illustrate the present invention and are not intended to limit the same.

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EXAMPLES**EXAMPLE 1: Synthesis of Oligonucleotides**

Unmodified oligodeoxynucleotides are synthesized on an automated DNA synthesizer (Applied Biosystems model 380B) using standard phosphoramidite chemistry with oxidation by iodine. β -cyanoethyl-diisopropyl-phosphoramidites are purchased from Applied Biosystems (Foster City, CA). For phosphorothioate oligonucleotides, the standard oxidation bottle was replaced by a 0.2 M solution of ^3H -1,2-benzodithiole-3-one 1,1-dioxide in acetonitrile for the stepwise thiation of the phosphite linkages. The thiation cycle wait step was increased to 68 seconds and was followed by the capping step. Cytosines may be 5-methyl cytosines. (5-methyl deoxycytidine phosphoramidites available from Glen Research, Sterling, VA or Amersham Pharmacia Biotech, Piscataway, NJ)

2'-methoxy oligonucleotides are synthesized using 2'-methoxy β -cyanoethyl-diisopropyl-phosphoramidites (Chemgenes, Needham, MA) and the standard cycle for unmodified oligonucleotides, except the wait step after pulse delivery of tetrazole and base is increased to 360 seconds. Other 2'-alkoxy oligonucleotides are synthesized by a modification of this method, using appropriate 2'-modified amidites such as those available from Glen Research, Inc., Sterling, VA.

2'-fluoro oligonucleotides are synthesized as described in Kawasaki et al. (*J. Med. Chem.* 1993, 36, 831-841). Briefly, the protected nucleoside N^6 -benzoyl-2'-deoxy-2'-fluoroadenosine is synthesized utilizing commercially available 9- β -D-arabinofuranosyladenine as starting material and by modifying literature procedures whereby the 2'- α -fluoro atom is introduced by a $\text{S}_{\text{N}}2$ -displacement of a 2'- β -O-triflyl group. Thus N^6 -benzoyl-9- β -D-arabinofuranosyladenine is selectively protected in moderate yield as the 3',5'-ditetrahydropyranyl (THP)

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intermediate. Deprotection of the THP and N⁶-benzoyl groups is accomplished using standard methodologies and standard methods are used to obtain the 5'-dimethoxytrityl- (DMT) and 5'-DMT-3'-phosphoramidite intermediates.

5 The synthesis of 2'-deoxy-2'-fluoroguanosine is accomplished using tetraisopropylidisiloxanyl (TPDS) protected 9-β-D-arabinofuranosylguanine as starting material, and conversion to the intermediate diisobutyryl-arabinofuranosylguanosine. Deprotection of the TPDS group
10 is followed by protection of the hydroxyl group with THP to give diisobutyryl di-THP protected arabinofuranosylguanine. Selective O-deacylation and triflation is followed by treatment of the crude product with fluoride, then deprotection of the THP groups. Standard methodologies are
15 used to obtain the 5'-DMT- and 5'-DMT-3'-phosphoramidites.

Synthesis of 2'-deoxy-2'-fluorouridine is accomplished by the modification of a known procedure in which 2, 2'-anhydro-1-β-D-arabinofuranosyluracil is treated with 70% hydrogen fluoride-pyridine. Standard procedures are used
20 to obtain the 5'-DMT and 5'-DMT-3'phosphoramidites.

2'-deoxy-2'-fluorocytidine is synthesized via amination of 2'-deoxy-2'-fluorouridine, followed by selective protection to give N⁴-benzoyl-2'-deoxy-2'-fluorocytidine. Standard procedures are used to obtain the
25 5'-DMT and 5'-DMT-3'phosphoramidites.

2'-(2-methoxyethyl)-modified amidites were synthesized according to Martin, P. (*Helv. Chim. Acta* 1995, 78, 486-506). For ease of synthesis, the last nucleotide may be a deoxynucleotide. 2'-O-CH₂CH₂OCH₃ cytosines may be 5-methyl
30 cytosines.

Synthesis of 5-Methyl cytosine monomers:

2,2'-Anhydro[1-(β-D-arabinofuranosyl)-5-methyluridine]:

5-Methyluridine (ribosylthymine, commercially available through Yamasa, Choshi, Japan) (72.0 g, 0.279 M),
35 diphenylcarbonate (90.0 g, 0.420 M) and sodium bicarbonate

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(2.0 g, 0.024 M) were added to DMF (300 mL). The mixture was heated to reflux, with stirring, allowing the evolved carbon dioxide gas to be released in a controlled manner. After 1 hour, the slightly darkened solution was concentrated under reduced pressure. The resulting syrup was poured into diethylether (2.5 L), with stirring. The product formed a gum. The ether was decanted and the residue was dissolved in a minimum amount of methanol (ca. 400 mL). The solution was poured into fresh ether (2.5 L) to yield a stiff gum. The ether was decanted and the gum was dried in a vacuum oven (60°C at 1 mm Hg for 24 h) to give a solid which was crushed to a light tan powder (57 g, 85% crude yield). The material was used as is for further reactions.

15 2'-O-Methoxyethyl-5-methyluridine:

2,2'-Anhydro-5-methyluridine (195 g, 0.81 M), tris(2-methoxyethyl)borate (231 g, 0.98 M) and 2-methoxyethanol (1.2 L) were added to a 2 L stainless steel pressure vessel and placed in a pre-heated oil bath at 160°C. After heating for 48 hours at 155-160°C, the vessel was opened and the solution evaporated to dryness and triturated with MeOH (200 mL). The residue was suspended in hot acetone (1 L). The insoluble salts were filtered, washed with acetone (150 mL) and the filtrate evaporated. The residue (280 g) was dissolved in CH₃CN (600 mL) and evaporated. A silica gel column (3 kg) was packed in CH₂Cl₂/acetone/MeOH (20:5:3) containing 0.5% Et₃NH. The residue was dissolved in CH₂Cl₂ (250 mL) and adsorbed onto silica (150 g) prior to loading onto the column. The product was eluted with the packing solvent to give 160 g (63%) of product.

20 2'-O-Methoxyethyl-5'-O-dimethoxytrityl-5-methyluridine:

2'-O-Methoxyethyl-5-methyluridine (160 g, 0.506 M) was co-evaporated with pyridine (250 mL) and the dried residue dissolved in pyridine (1.3 L). A first aliquot of di-methoxytrityl chloride (94.3 g, 0.278 M) was added and the

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mixture stirred at room temperature for one hour. A second aliquot of dimethoxytrityl chloride (94.3 g, 0.278 M) was added and the reaction stirred for an additional one hour. Methanol (170 mL) was then added to stop the reaction.

5 HPLC showed the presence of approximately 70% product. The solvent was evaporated and triturated with CH₃CN (200 mL). The residue was dissolved in CHCl₃ (1.5 L) and extracted with 2x500 mL of saturated NaHCO₃ and 2x500 mL of saturated NaCl. The organic phase was dried over Na₂SO₄, filtered and
10 evaporated. 275 g of residue was obtained. The residue was purified on a 3.5 kg silica gel column, packed and eluted with EtOAc/Hexane/Acetone (5:5:1) containing 0.5% Et₃NH. The pure fractions were evaporated to give 164 g of product. Approximately 20 g additional was obtained from
15 the impure fractions to give a total yield of 183 g (57%).

3'-O-Acetyl-2'-O-methoxyethyl-5'-O-dimethoxytrityl-5-methyluridine:

2'-O-Methoxyethyl-5'-O-dimethoxytrityl-5-methyluridine (106 g, 0.167 M), DMF/pyridine (750 mL of a 3:1 mixture
20 prepared from 562 mL of DMF and 188 mL of pyridine) and acetic anhydride (24.38 mL, 0.258 M) were combined and stirred at room temperature for 24 hours. The reaction was monitored by tlc by first quenching the tlc sample with the addition of MeOH. Upon completion of the reaction, as
25 judged by tlc, MeOH (50 mL) was added and the mixture evaporated at 35°C. The residue was dissolved in CHCl₃ (800 mL) and extracted with 2x200 mL of saturated sodium bicarbonate and 2x200 mL of saturated NaCl. The water layers were back extracted with 200 mL of CHCl₃. The
30 combined organics were dried with sodium sulfate and evaporated to give 122 g of residue (approx. 90% product). The residue was purified on a 3.5 kg silica gel column and eluted using EtOAc/Hexane(4:1). Pure product fractions were evaporated to yield 96 g (84%).

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3'-O-Acetyl-2'-O-methoxyethyl-5'-O-dimethoxytrityl-5-methyl-4-triazoleuridine:

A first solution was prepared by dissolving 3'-O-acetyl-2'-O-methoxyethyl-5'-O-dimethoxytrityl-5-methyluridine (96 g, 0.144 M) in CH₃CN (700 mL) and set aside. Triethylamine (189 mL, 1.44 M) was added to a solution of triazole (90 g, 1.3 M) in CH₃CN (1 L), cooled to -5°C and stirred for 0.5 h using an overhead stirrer. POCl₃ was added dropwise, over a 30 minute period, to the stirred solution maintained at 0-10°C, and the resulting mixture stirred for an additional 2 hours. The first solution was added dropwise, over a 45 minute period, to the later solution. The resulting reaction mixture was stored overnight in a cold room. Salts were filtered from the reaction mixture and the solution was evaporated. The residue was dissolved in EtOAc (1 L) and the insoluble solids were removed by filtration. The filtrate was washed with 1x300 mL of NaHCO₃ and 2x300 mL of saturated NaCl, dried over sodium sulfate and evaporated. The residue was triturated with EtOAc to give the title compound.

2'-O-Methoxyethyl-5'-O-dimethoxytrityl-5-methylcytidine:

A solution of 3'-O-acetyl-2'-O-methoxyethyl-5'-O-dimethoxytrityl-5-methyl-4-triazoleuridine (103 g, 0.141 M) in dioxane (500 mL) and NH₄OH (30 mL) was stirred at room temperature for 2 hours. The dioxane solution was evaporated and the residue azeotroped with MeOH (2x200 mL). The residue was dissolved in MeOH (300 mL) and transferred to a 2 liter stainless steel pressure vessel. MeOH (400 mL) saturated with NH₃ gas was added and the vessel heated to 100°C for 2 hours (tlc showed complete conversion). The vessel contents were evaporated to dryness and the residue was dissolved in EtOAc (500 mL) and washed once with saturated NaCl (200 mL). The organics were dried over sodium sulfate and the solvent was evaporated to give 85 g (95%) of the title compound.

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N⁴-Benzoyl-2'-O-methoxyethyl-5'-O-dimethoxytrityl-5-methylcytidine:

2'-O-Methoxyethyl-5'-O-dimethoxytrityl-5-methylcytidine (85 g, 0.134 M) was dissolved in DMF (800 mL) and 5 benzoic anhydride (37.2 g, 0.165 M) was added with stirring. After stirring for 3 hours, tlc showed the reaction to be approximately 95% complete. The solvent was evaporated and the residue azeotroped with MeOH (200 mL). The residue was dissolved in CHCl₃ (700 mL) and extracted 10 with saturated NaHCO₃ (2x300 mL) and saturated NaCl (2x300 mL), dried over MgSO₄ and evaporated to give a residue (96 g). The residue was chromatographed on a 1.5 kg silica column using EtOAc/Hexane (1:1) containing 0.5% Et₃NH as the eluting solvent. The pure product fractions were 15 evaporated to give 90 g (90%) of the title compound.

N⁴-Benzoyl-2'-O-methoxyethyl-5'-O-dimethoxytrityl-5-methylcytidine-3'-amidite:

N⁴-Benzoyl-2'-O-methoxyethyl-5'-O-dimethoxytrityl-5-methylcytidine (74 g, 0.10 M) was dissolved in CH₂Cl₂ (1 20 L). Tetrazole diisopropylamine (7.1 g) and 2-cyanoethoxytetra(isopropyl)phosphite (40.5 mL, 0.123 M) were added with stirring, under a nitrogen atmosphere. The resulting mixture was stirred for 20 hours at room temperature (tlc showed the reaction to be 95% complete). The reaction 25 mixture was extracted with saturated NaHCO₃ (1x300 mL) and saturated NaCl (3x300 mL). The aqueous washes were back-extracted with CH₂Cl₂ (300 mL), and the extracts were combined, dried over MgSO₄ and concentrated. The residue obtained was chromatographed on a 1.5 kg silica column 30 using EtOAc/Hexane (3:1) as the eluting solvent. The pure fractions were combined to give 90.6 g (87%) of the title compound.

5-methyl-2'-deoxycytidine (5-me-C) containing oligonucleotides were synthesized according to published 35 methods (Sanghvi et al., Nucl. Acids Res. 1993, 21, 3197-

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3203) using commercially available phosphoramidites (Glen Research, Sterling VA or ChemGenes, Needham MA).

2'-O-(dimethylaminooxyethyl) nucleoside amidites

2'-(Dimethylaminooxyethoxy) nucleoside amidites [also
5 known in the art as 2'-O-(dimethylaminooxyethyl) nucleoside
amidites] are prepared as described in the following
paragraphs. Adenosine, cytidine and guanosine nucleoside
amidites are prepared similarly to the thymidine (5-
methyluridine) except the exocyclic amines are protected
10 with a benzoyl moiety in the case of adenosine and cytidine
and with isobutyryl in the case of guanosine.

5'-O-tert-Butyldiphenylsilyl-O²-2'-anhydro-5-methyluridine

O²-2'-anhydro-5-methyluridine (Pro. Bio. Sint., Varese,
Italy, 100.0g, 0.416 mmol), dimethylaminopyridine (0.66g,
15 0.013eq, 0.0054mmol) were dissolved in dry pyridine (500
ml) at ambient temperature under an argon atmosphere and
with mechanical stirring. tert-Butyldiphenylchlorosilane
(125.8g, 119.0mL, 1.1eq, 0.458mmol) was added in one
portion. The reaction was stirred for 16 h at ambient
20 temperature. TLC (R_f 0.22, ethyl acetate) indicated a
complete reaction. The solution was concentrated under
reduced pressure to a thick oil. This was partitioned
between dichloromethane (1 L) and saturated sodium
bicarbonate (2x1 L) and brine (1 L). The organic layer was
25 dried over sodium sulfate and concentrated under reduced
pressure to a thick oil. The oil was dissolved in a 1:1
mixture of ethyl acetate and ethyl ether (600mL) and the
solution was cooled to -10°C. The resulting crystalline
product was collected by filtration, washed with ethyl
30 ether (3x200 mL) and dried (40°C, 1mm Hg, 24 h) to 149g
(74.8%) of white solid. TLC and NMR were consistent with
pure product.

5'-O-tert-Butyldiphenylsilyl-2'-O-(2-hydroxyethyl)-5-
methyluridine

35 In a 2 L stainless steel, unstirred pressure reactor

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was added borane in tetrahydrofuran (1.0 M, 2.0 eq, 622 mL). In the fume hood and with manual stirring, ethylene glycol (350 mL, excess) was added cautiously at first until the evolution of hydrogen gas subsided. 5'-O-tert-Butyldiphenylsilyl-O²-2'-anhydro-5-methyluridine (149 g, 0.311 mol) and sodium bicarbonate (0.074 g, 0.003 eq) were added with manual stirring. The reactor was sealed and heated in an oil bath until an internal temperature of 160 °C was reached and then maintained for 16 h (pressure < 100 psig). The reaction vessel was cooled to ambient and opened. TLC (R_f 0.67 for desired product and R_f 0.82 for ara-T side product, ethyl acetate) indicated about 70% conversion to the product. In order to avoid additional side product formation, the reaction was stopped, concentrated under reduced pressure (10 to 1mm Hg) in a warm water bath (40-100°C) with the more extreme conditions used to remove the ethylene glycol. [Alternatively, once the low boiling solvent is gone, the remaining solution can be partitioned between ethyl acetate and water. The product will be in the organic phase.] The residue was purified by column chromatography (2kg silica gel, ethyl acetate-hexanes gradient 1:1 to 4:1). The appropriate fractions were combined, stripped and dried to product as a white crisp foam (84g, 50%), contaminated starting material (17.4g) and pure reusable starting material 20g. The yield based on starting material less pure recovered starting material was 58%. TLC and NMR were consistent with 99% pure product.

30 2'-O-([2-phthalimidoxy)ethyl]-5'-t-butyldiphenylsilyl-5-methyluridine

5'-O-tert-Butyldiphenylsilyl-2'-O-(2-hydroxyethyl)-5-methyluridine (20g, 36.98mmol) was mixed with triphenylphosphine (11.63g, 44.36mmol) and N-hydroxyphthalimide (7.24g, 44.36mmol). It was then dried over P₂O₅ under high vacuum for two days at 40°C. The

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reaction mixture was flushed with argon and dry THF (369.8mL, Aldrich, sure seal bottle) was added to get a clear solution. Diethyl-azodicarboxylate (6.98mL, 44.36mmol) was added dropwise to the reaction mixture. The rate of addition is maintained such that resulting deep red coloration is just discharged before adding the next drop. After the addition was complete, the reaction was stirred for 4 hrs. By that time TLC showed the completion of the reaction (ethylacetate:hexane, 60:40). The solvent was evaporated in vacuum. Residue obtained was placed on a flash column and eluted with ethyl acetate:hexane (60:40), to get 2'-O-([2-phthalimidoxy)ethyl]-5'-t-butyl-
10 butyldiphenylsilyl-5-methyluridine as white foam (21.819, 86%).

15 5'-O-tert-butyldiphenylsilyl-2'-O-[(2-formadoximinooxy)ethyl]-5-methyluridine

2'-O-([2-phthalimidoxy)ethyl]-5'-t-butyl-
5-methyluridine (3.1g, 4.5mmol) was dissolved in dry CH₂Cl₂ (4.5mL) and methylhydrazine (300mL, 4.64mmol) was added dropwise at -10°C to 0°C. After 1 hr the mixture was
20 filtered, the filtrate was washed with ice cold CH₂Cl₂ and the combined organic phase was washed with water, brine and dried over anhydrous Na₂SO₄. The solution was concentrated to get 2'-O-(aminooxyethyl) thymidine, which was then
25 dissolved in MeOH (67.5mL). To this formaldehyde (20% aqueous solution, w/w, 1.1eg.) was added and the mixture for 1 hr. Solvent was removed under vacuum; residue chromatographed to get 5'-O-tert-butyldiphenylsilyl-2'-O-
[(2-formadoximinooxy) ethyl]-5-methyluridine as white foam
30 (1.95, 78%).

5'-O-tert-Butyldiphenylsilyl-2'-O-[N,N-dimethylaminooxyethyl]-5-methyluridine

5'-O-tert-butyldiphenylsilyl-2'-O-[(2-formadoximinooxy)ethyl]-5-methyluridine (1.77g, 3.12mmol)
35 was dissolved in a solution of 1M pyridinium p-

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toluenesulfonate (PPTS) in dry MeOH (30.6mL). Sodium cyanoborohydride (0.39g, 6.13mmol) was added to this solution at 10°C under inert atmosphere. The reaction mixture was stirred for 10 minutes at 10°C. After that the reaction vessel was removed from the ice bath and stirred at room temperature for 2 hr, the reaction monitored by TLC (5% MeOH in CH₂Cl₂). Aqueous NaHCO₃ solution (5%, 10mL) was added and extracted with ethyl acetate (2x20mL). Ethyl acetate phase was dried over anhydrous Na₂SO₄, evaporated to dryness. Residue was dissolved in a solution of 1M PPTS in MeOH (30.6mL). Formaldehyde (20% w/w, 30mL, 3.37mmol) was added and the reaction mixture was stirred at room temperature for 10 minutes. Reaction mixture cooled to 10°C in an ice bath, sodium cyanoborohydride (0.39g, 6.13mmol) was added and reaction mixture stirred at 10°C for 10 minutes. After 10 minutes, the reaction mixture was removed from the ice bath and stirred at room temperature for 2 hrs. To the reaction mixture 5% NaHCO₃ (25mL) solution was added and extracted with ethyl acetate (2x25mL). Ethyl acetate layer was dried over anhydrous Na₂SO₄ and evaporated to dryness. The residue obtained was purified by flash column chromatography and eluted with 5% MeOH in CH₂Cl₂ to get 5'-O-tert-butyldiphenylsilyl-2'-O-[N,N-dimethylaminooxyethyl]-5-methyluridine as a white foam (14.6g, 80%).

2'-O-(dimethylaminoxyethyl)-5-methyluridine

Triethylamine trihydrofluoride (3.91mL, 24.0mmol) was dissolved in dry THF and triethylamine (1.67mL, 12mmol, dry, kept over KOH). This mixture of triethylamine-2HF was then added to 5'-O-tert-butyldiphenylsilyl-2'-O-[N,N-dimethylaminoxyethyl]-5-methyluridine (1.40g, 2.4mmol) and stirred at room temperature for 24 hrs. Reaction was monitored by TLC (5% MeOH in CH₂Cl₂). Solvent was removed under vacuum and the residue placed on a flash column and eluted with 10% MeOH in CH₂Cl₂ to get 2'-O-

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(dimethylaminoxyethyl)-5-methyluridine (766mg, 92.5%).
5'-O-DMT-2'-O-(dimethylaminoxyethyl)-5-methyluridine
2'-O-(dimethylaminoxyethyl)-5-methyluridine (750mg,
2.17mmol) was dried over P_2O_5 under high vacuum overnight at
5 40°C. It was then co-evaporated with anhydrous pyridine
(20mL). The residue obtained was dissolved in pyridine
(11mL) under argon atmosphere. 4-dimethylaminopyridine
(26.5mg, 2.60mmol), 4,4'-dimethoxytrityl chloride (880mg,
2.60mmol) was added to the mixture and the reaction mixture
10 was stirred at room temperature until all of the starting
material disappeared. Pyridine was removed under vacuum
and the residue chromatographed and eluted with 10% MeOH in
 CH_2Cl_2 (containing a few drops of pyridine) to get 5'-O-DMT-
2'-O-(dimethylamino-oxyethyl)-5-methyluridine (1.13g, 80%).
15 5'-O-DMT-2'-O-(2-N,N-dimethylaminoxyethyl)-5-
methyluridine-3'-[(2-cyanoethyl)-N,N-
diisopropylphosphoramidite]
5'-O-DMT-2'-O-(dimethylaminoxyethyl)-5-methyluridine
(1.08g, 1.67mmol) was co-evaporated with toluene (20mL).
20 To the residue N,N-diisopropylamine tetrazonide (0.29g,
1.67mmol) was added and dried over P_2O_5 under high vacuum
overnight at 40°C. Then the reaction mixture was dissolved
in anhydrous acetonitrile (8.4mL) and 2-cyanoethyl-
N,N,N¹,N¹-tetraisopropylphosphoramidite (2.12mL, 6.08mmol)
25 was added. The reaction mixture was stirred at ambient
temperature for 4 hrs under inert atmosphere. The progress
of the reaction was monitored by TLC (hexane:ethyl acetate
1:1). The solvent was evaporated, then the residue was
dissolved in ethyl acetate (70mL) and washed with 5%
30 aqueous $NaHCO_3$ (40mL). Ethyl acetate layer was dried over
anhydrous Na_2SO_4 and concentrated. Residue obtained was
chromatographed (ethyl acetate as eluent) to get 5'-O-DMT-
2'-O-(2-N,N-dimethylaminoxyethyl)-5-methyluridine-3'-[(2-
cyanoethyl)-N,N-diisopropylphosphoramidite] as a foam
35 (1.04g, 74.9%).

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Oligonucleotides having methylene(methylimino) (MMI) backbones are synthesized according to U.S. Patent 5,378,825, which is coassigned to the assignee of the present invention and is incorporated herein in its entirety. For ease of synthesis, various nucleoside dimers containing MMI linkages are synthesized and incorporated into oligonucleotides. Other nitrogen-containing backbones are synthesized according to WO 92/20823 which is also coassigned to the assignee of the present invention and incorporated herein in its entirety.

Oligonucleotides having amide backbones are synthesized according to De Mesmaeker et al. (*Acc. Chem. Res.* 1995, 28, 366-374). The amide moiety is readily accessible by simple and well-known synthetic methods and is compatible with the conditions required for solid phase synthesis of oligonucleotides.

Oligonucleotides with morpholino backbones are synthesized according to U.S. Patent 5,034,506 (Summerton and Weller).

Peptide-nucleic acid (PNA) oligomers are synthesized according to P.E. Nielsen et al. (*Science* 1991, 254, 1497-1500).

After cleavage from the controlled pore glass column (Applied Biosystems) and deblocking in concentrated ammonium hydroxide at 55°C for 18 hours, the oligonucleotides are purified by precipitation twice out of 0.5 M NaCl with 2.5 volumes ethanol. Synthesized oligonucleotides were analyzed by polyacrylamide gel electrophoresis on denaturing gels or capillary gel electrophoresis and judged to be at least 85% full length material. The relative amounts of phosphorothioate and phosphodiester linkages obtained in synthesis were periodically checked by ³¹P nuclear magnetic resonance spectroscopy, and for some studies oligonucleotides were purified by HPLC, as described by Chiang et al. (*J. Biol.*

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Chem. 1991, 266, 18162). Results obtained with HPLC-purified material were similar to those obtained with non-HPLC purified material.

Alternatively, oligonucleotides were synthesized in 96 well plate format via solid phase P(III) phosphoramidite chemistry on an automated synthesizer capable of assembling 96 sequences simultaneously in a standard 96 well format. Phosphodiester internucleotide linkages were afforded by oxidation with aqueous iodine. Phosphorothioate internucleotide linkages were generated by sulfurization utilizing 3,4-dithiolane-2-one 1,1-dioxide (Beaucage Reagent) in anhydrous acetonitrile. Standard base-protected beta-cyanoethyl-di-isopropyl phosphoramidites were purchased from commercial vendors (e.g. PE-Applied Biosystems, Foster City, CA, or Pharmacia, Piscataway, NJ). Non-standard nucleosides are synthesized as per published methods. They are utilized as base protected beta-cyanoethyl-diisopropyl phosphoramidites.

Oligonucleotides were cleaved from support and deprotected with concentrated NH_4OH at elevated temperature (55-60°C) for 12-16 hours and the released product then dried in vacuo. The dried product was then re-suspended in sterile water to afford a master plate from which all analytical and test plate samples are then diluted utilizing robotic pipettors.

EXAMPLE 2: Human STAT3 Oligodeoxynucleotide Sequences

Antisense oligonucleotides were designed to target human STAT3. Target sequence data are from the APRF cDNA sequence published by Akira, S. et al. (Cell, 1994, 77, 63-71); Genbank accession number L29277, provided herein as SEQ ID NO: 1. A set of oligodeoxynucleotides were synthesized with phosphorothioate linkages. 2'-deoxy cytosines were 5-methyl cytosines. These oligonucleotide sequences are shown in Table 1. An additional set of oligonucleotides was synthesized as chimeric

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oligonucleotides ("gapmers") 20 nucleotides in length, composed of a central "gap" region consisting of ten 2'-deoxynucleotides, which is flanked on both sides (5' and 3' directions) by five-nucleotide "wings." The wings are composed of 2'-methoxyethyl (2'-MOE)nucleotides. The internucleoside (backbone) linkages are phosphorothioate (P=S) throughout the oligonucleotide. All 2'-MOE cytosines and 2'-deoxy cytosines were 5-methyl-cytosines. These oligonucleotide sequences are shown in Table 2.

An appropriate cell line, typically expressing high levels of STAT3, is chosen for *in vitro* studies. Cell culture conditions are those standard for that particular cell line. Oligonucleotide treatment is for four hours and mRNA usually isolated 24 to 48 hours following initial treatment. mRNA is isolated using the RNeasy® kit (Qiagen, Santa Clarita, CA).

TABLE 1:
Nucleotide Sequences of Human STAT3 Phosphorothioate Oligodeoxynucleotides

ISIS NO.	NUCLEOTIDE SEQUENCE ¹ (5' -> 3')	SEQ ID NO:	TARGET GENE NUCLEOTIDE CO-ORDINATES ²	GENE TARGET REGION
106691	GTCTGCGCCGCGCCCCGAA	2	0010-0029	5'-UTR
106692	GGCCGAAGGGCCTCTCCGAG	3	0130-0149	5'-UTR
106693	TCCTGTTTCTCCGGCAGAGG	4	0202-0221	AUG
106694	CATCCTGTTTCTCCGGCAGA	5	0204-0223	AUG
106695	GCCATCCTGTTTCTCCGGCA	6	0206-0225	AUG
106696	GGGCCATCCTGTTTCTCCGG	7	0208-0227	AUG
106697	TTGGGCCATCCTGTTTCTCC	8	0210-0229	AUG
106698	CATTGGGCCATCCTGTTTCT	9	0212-0231	AUG
106699	TCCATTGGGCCATCCTGTTT	10	0214-0233	AUG
106700	ATTCCATTGGGCCATCCTGT	11	0216-0235	AUG
106701	TGATTCCATTGGGCCATCCT	12	0218-0237	AUG

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	106702	GCTGATTCCATTGGGCCATC	13	0220-0239	AUG
	106703	TAGCTGATTCCATTGGGCCA	14	0222-0241	AUG
	106704	TGTAGCTGATTCCATTGGGC	15	0224-0243	coding
	106705	CTGTAGAGCTGATGGAGCTG	16	0269-0288	coding
5	106706	CCCAATCTTGACTCTCAATC	17	0331-0350	coding
	106707	CCCAGGAGATTATGAAACAC	18	0386-0405	coding
	106708	ACATTCGACTCTTGACAGGAA	19	0431-0450	coding
	106709	TCTGAAGAACTGCTTGATT	20	0475-0494	coding
	106710	GGCCACAATCCGGGCAATCT	21	0519-0538	coding
10	106711	TGGCTGCAGTCTGTAGAAGG	22	0562-0581	coding
	106712	CTGCTCCAGCATCTGCTGCT	23	0639-0658	coding
	106713	TTTCTGTTCTAGATCCTGCA	24	0684-0703	coding
	106714	TAGTTGAAATCAAAGTCATC	25	0728-0747	coding
	106715	TTCCATTCAGATCTTGCATG	26	0772-0791	coding
15	106716	TCTGTTCCAGCTGCTGCATC	27	0817-0836	coding
	106717	TCACTCACGATGCTTCTCCG	28	0860-0879	coding
	106718	GAGTTTTCTGCACGTACTCC	29	0904-0923	coding
	106719	ATCTGTTGCCGCCTCTTCCA	30	0947-0968	coding
	106720	CTAGCCGATCTAGGCAGATG	31	0991-1010	coding
20	106721	CGGGTCTGAAGTTGAGATTC	32	1034-1053	coding
	106722	CGGCCGGTGCTGTACAATGG	33	1110-1129	coding
	106723	TTTCATTAAGTTTCTGAACA	34	1155-1174	coding
	106724	AGGATGCATGGGCATGCAGG	35	1200-1219	coding
	106725	GACCAGCAACCTGACTTTAG	36	1260-1279	coding
25	106726	ATGCACACTTTAATTTTAAG	37	1304-1323	coding
	106727	TTCCGGGATCCTCTGAGAGC	38	1349-1368	coding
	106728	TTCCATGTTCATCACTTTTG	39	1392-1411	coding
	106729	GTCAAGTGTTTGAATTCTGC	40	1436-1455	coding
	106730	CAATCAGGGAAGCATCACAA	41	1495-1514	coding
30	106731	TACACCTCGGTCTCAAAGGT	42	1538-1557	coding

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	106732	TGACAAGGAGTGGGTCTCTA	43	1581-1600	coding
	106733	CGCCCAGGCATTTGGCATCT	44	1626-1645	coding
	106734	CATTCTTGGGATTGTTGGTC	45	1669-1688	coding
	106735	CACTTGGTCCCAGGTTCCAA	46	1713-1732	coding
5	106736	CCCGCTTGGTGGTGGACGAG	47	1756-1775	coding
	106737	AGTTCACACCAGGCCCTAGG	48	1816-1835	coding
	106738	GTTTTCTTTGCAGAAGTTAG	49	1860-1879	coding
	106739	ATATTGTCTAGCCAGACCCA	50	1904-1923	coding
	106740	AACCCATGATGTACCCTTCA	51	1963-1982	coding
10	106741	GCTTAGTGCTCAAGATGGCC	52	2005-2024	coding
	106742	GCTGCTTTCACTGAAGCGCA	53	2043-2062	coding
	106743	GTGAAAGTGACGCCTCCTTC	54	2066-2085	coding
	106744	CTGATGTCCTTCTCCACCCA	55	2087-2106	coding
	106745	ACTGGATCTGGGTCTTACCG	56	2107-2126	coding
15	106746	AAATGACATGTTGTTTCAGCT	57	2151-2170	coding
	106747	GCCCATGATGATTTTCAGCAA	58	2169-2188	coding
	106748	TATTGGTAGCATCCATGATC	59	2194-2213	coding
	106749	ATAGACAAGTGGAGACAACA	60	2217-2236	coding
	106750	TTGGGAATGTCAGGATAGAG	61	2237-2256	coding
20	106751	CTCCTGGCTCTCTGGCCGAC	62	2280-2299	coding
	106752	ACCTGGGTCAGCTTCAGGAT	63	2301-2320	coding
	106753	CACAGATAAACTTGGTCTTC	64	2338-2357	coding
	106754	ATCGGCAGGTCAATGGTATT	65	2378-2397	coding
	106755	CCAAACTGCATCAATGAATC	66	2414-2433	coding
25	106756	GGTTCAGCACCTTCACCATT	67	2438-2457	coding
	106757	GAGGGACTCAAACCTGCCCTC	68	2466-2485	coding
	106758	CAACTCCATGTCAAAGGTGA	69	2484-2503	coding
	106759	TTCTCAGCTCCTCACATGGG	70	2525-2544	STOP
	106760	CGTTCTCAGCTCCTCACATG	71	2527-2546	STOP
30	106761	TCCGTTCTCAGCTCCTCACA	72	2529-2548	STOP

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	106762	CTTCCGTTCTCAGCTCCTCA	73	2531-2550	STOP
	106763	AGCTTCCGTTCTCAGCTCCT	74	2533-2552	STOP
	106764	AGAATGCAGGTAGGCGCCTC	75	2569-2588	3'-UTR
	106765	ACCACAAAGTTAGTAGTTTC	76	2623-2642	3'-UTR
5	106766	TGCTCAAAGATAGCAGAAGT	77	2665-2684	3'-UTR
	106767	ATTCACTCATTTCTCTATTT	78	2701-2720	3'-UTR
	106768	CATTTAGATAAAAGCAGATC	79	2727-2746	3'-UTR
	106769	ACATCCTTATTTGCATTTAG	80	2740-2759	3'-UTR
	106770	GATCATGGGTCTCAGAGAAC	81	2760-2779	3'-UTR

10

¹ "C" residues are 5-methyl-cytosines; all linkages are phosphorothioate linkages.

² Coordinates from Genbank Accession No. L29277, locus name "HUMAPRF", SEQ ID NO. 1.

15

TABLE 2:

Nucleotide Sequences of Human STAT3 Chimeric (deoxy gapped)
Phosphorothioate Oligonucleotides

	ISIS NO.	NUCLEOTIDE SEQUENCE ¹ (5' -> 3')	SEQ ID NO:	TARGET GENE NUCLEOTIDE CO-ORDINATES ²	GENE TARGET REGION
20	106771	GTCTGCGCCGCGCCCCGAA	2	0010-0029	5'-UTR
	106772	GGCCGAAGGGCCTCTCCGAG	3	0130-0149	5'-UTR
	106773	TCCTGTTTCTCCGGCAGAGG	4	0202-0221	AUG
	106774	CATCCTGTTTCTCCGGCAGA	5	0204-0223	AUG
25	106775	GCCATCCTGTTTCTCCGGCA	6	0206-0225	AUG
	106776	GGGCCATCCTGTTTCTCCGG	7	0208-0227	AUG
	106777	TTGGGCCATCCTGTTTCTCC	8	0210-0229	AUG
	106778	CATTGGGCCATCCTGTTTCT	9	0212-0231	AUG
	106779	TCCATTGGGCCATCCTGTTT	10	0214-0233	AUG
30	106780	ATTCCATTGGGCCATCCTGT	11	0216-0235	AUG
	106781	TGATTCCATTGGGCCATCCT	12	0218-0237	AUG
	106782	GCTGATTCCATTGGGCCATC	13	0220-0239	AUG

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	106783	TAGCTGATTCCATTGGGCCA	14	0222-0241	AUG
	106784	TGTAGCTGATTCCATTGGGC	15	0224-0243	coding
	106785	CTGTAGAGCTGATGGAGCTG	16	0269-0288	coding
	106786	CCCAATCTTGA CTCTCAATC	17	0331-0350	coding
5	106787	CCCAGGAGATTATGAAACAC	18	0386-0405	coding
	106788	ACATTGACTCTTGCAGGAA	19	0431-0450	coding
	106789	TCTGAAGAACTGCTTGATT	20	0475-0494	coding
	106790	GGCCACAATCCGGGCAATCT	21	0519-0538	coding
	106791	TGGCTGCAGTCTGTAGAAGG	22	0562-0581	coding
10	106792	CTGCTCCAGCATCTGCTGCT	23	0639-0658	coding
	106793	TTTCTGTTCTAGATCCTGCA	24	0684-0703	coding
	106794	TAGTTGAAATCAAAGTCATC	25	0728-0747	coding
	106795	TTCCATTCAGATCTTG CATG	26	0772-0791	coding
	106796	TCTGTTCCAGCTGCTGCATC	27	0817-0836	coding
15	106797	TCACTCACGATGCTTCTCCG	28	0860-0879	coding
	106798	GAGTTTTCTGCACGTACTCC	29	0904-0923	coding
	106799	ATCTGTTGCCGCCTCTTCCA	30	0947-0968	coding
	106800	CTAGCCGATCTAGGCAGATG	31	0991-1010	coding
	106801	CGGGTCTGAAGTTGAGATTC	32	1034-1053	coding
20	106802	CGGCCGGTGCTGTACAATGG	33	1110-1129	coding
	106803	TTTCATTAAGTTTCTGAACA	34	1155-1174	coding
	106804	AGGATGCATGGGCATGCAGG	35	1200-1219	coding
	106805	GACCAGCAACCTGACTTTAG	36	1260-1279	coding
	106806	ATGCACACTTTAATTTTAAG	37	1304-1323	coding
25	106807	TTCCGGGATCCTCTGAGAGC	38	1349-1368	coding
	106808	TTCCATGTT CATCACTTTTG	39	1392-1411	coding
	106809	GTCAAGTGTTTGAATTCTGC	40	1436-1455	coding
	106810	CAATCAGGGAAGCATCACAA	41	1495-1514	coding
	106811	TACACCTCGGTCTCAAAGGT	42	1538-1557	coding

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	106812	TGACAAGGAGTGGGTCTCTA	43	1581-1600	coding
	106813	CGCCAGGCATTTGGCATCT	44	1626-1645	coding
	106814	CATTCTTGGGATTGTTGGTC	45	1669-1688	coding
	106815	CACCTTGGTCCCAGGTTCCTAA	46	1713-1732	coding
5	106816	CCCGCTTGGTGGTGGACGAG	47	1756-1775	coding
	106817	AGTTTACACACCACCCCTAGG	48	1816-1835	coding
	106818	GTTTTCTTTGCAGAAGTTAG	49	1860-1879	coding
	106819	ATATTGTCTAGCCAGACCCA	50	1904-1923	coding
	106820	AACCCATGATGTACCCTTCA	51	1963-1982	coding
10	106821	GCTTAGTGCTCAAGATGGCC	52	2005-2024	coding
	106822	GCTGCTTTCACTGAAGCGCA	53	2043-2062	coding
	106823	GTGAAAGTGACGCCTCCTTC	54	2066-2085	coding
	106824	CTGATGTCCTTCTCCACCCA	55	2087-2106	coding
	106825	ACTGGATCTGGGTCTTACCG	56	2107-2126	coding
15	106826	AAATGACATGTTGTTCAAGCT	57	2151-2170	coding
	106827	GCCCATGATGATTTTCAAGCA	58	2169-2188	coding
	106828	TATTGGTAGCATCCATGATC	59	2194-2213	coding
	106829	ATAGACAAGTGGAGACAACA	60	2217-2236	coding
	106830	TTGGGAATGTCAGGATAGAG	61	2237-2256	coding
20	106831	CTCCTGGCTCTCTGGCCGAC	62	2280-2299	coding
	106832	ACCTGGGTCAGCTTCAGGAT	63	2301-2320	coding
	106833	CACAGATAAACTTGGTCTTC	64	2338-2357	coding
	106834	ATCGGCAGGTCAATGGTATT	65	2378-2397	coding
	106835	CCAAACTGCATCAATGAATC	66	2414-2433	coding
25	106836	GGTTCAGCACCTTCACCATT	67	2438-2457	coding
	106837	GAGGGACTCAAAGTGCCTTC	68	2466-2485	coding
	106838	CAACTCCATGTCAAAGGTGA	69	2484-2503	coding
	106839	TTCTCAGCTCCTCACATGGG	70	2525-2544	STOP
	106840	CGTTCTCAGCTCCTCACATG	71	2527-2546	STOP

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	106841	TCCGTTCTCAGCTCCTCACA	72	2529-2548	STOP
	106842	CTTCCGTTCTCAGCTCCTCA	73	2531-2550	STOP
	106843	AGCTTCCGTTCTCAGCTCCT	74	2533-2552	STOP
	106844	AGAATGCAGGTAGGCGCCTC	75	2569-2588	3'-UTR
5	106845	ACCACAAAGTTAGTAGTTTC	76	2623-2642	3'-UTR
	106846	TGCTCAAAGATAGCAGAAGT	77	2665-2684	3'-UTR
	106847	ATTCACTCATTTCTCTATTT	78	2701-2720	3'-UTR
	106848	CATTTAGATAAAAGCAGATC	79	2727-2746	3'-UTR
	106849	ACATCCTTATTTGCATTTAG	80	2740-2759	3'-UTR
10	106850	GATCATGGGTCTCAGAGAAC	81	2760-2779	3'-UTR

¹ Emboldened residues are 2'-methoxyethoxy residues, 2'-methoxyethoxy cytosine residues and 2'-OH cytosine residues are 5-methyl-cytosines; all linkages are phosphorothioate linkages.

² Coordinates from Genbank Accession No. L29277, locus name "HUMAPRF", SEQ ID NO. 1.

Oligonucleotide activity is assayed by quantitation of STAT3 mRNA levels by real-time PCR (RT-PCR) using the ABI PRISM™ 7700 Sequence Detection System (PE-Applied Biosystems, Foster City, CA) according to manufacturer's instructions. This is a closed-tube, non-gel-based, fluorescence detection system which allows high-throughput quantitation of polymerase chain reaction (PCR) products in real-time. As opposed to standard PCR, in which amplification products are quantitated after the PCR is completed, products in RT-PCR are quantitated as they accumulate. This is accomplished by including in the PCR reaction an oligonucleotide probe that anneals specifically between the forward and reverse PCR primers, and contains two fluorescent dyes. A reporter dye (e.g., JOE or FAM, PE-Applied Biosystems, Foster City, CA) is attached to the

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5' end of the probe and a quencher dye (e.g., TAMRA, PE-Applied Biosystems, Foster City, CA) is attached to the 3' end of the probe. When the probe and dyes are intact, reporter dye emission is quenched by the proximity of the 3' quencher dye. During amplification, annealing of the probe to the target sequence creates a substrate that can be cleaved by the 5'-exonuclease activity of Taq polymerase. During the extension phase of the PCR amplification cycle, cleavage of the probe by Taq polymerase releases the reporter dye from the remainder of the probe (and hence from the quencher moiety) and a sequence-specific fluorescent signal is generated. With each cycle, additional reporter dye molecules are cleaved from their respective probes, and the fluorescence intensity is monitored at regular (six-second) intervals by laser optics built into the ABI PRISM™ 7700 Sequence Detection System. In each assay, a series of parallel reactions containing serial dilutions of mRNA from untreated control samples generates a standard curve that is used to quantitate the percent inhibition after antisense oligonucleotide treatment of test samples.

RT-PCR reagents are obtained from PE-Applied Biosystems, Foster City, CA. RT-PCR reactions are carried out by adding 25 µl PCR cocktail (1x TAQMAN® buffer A, 5.5 mM MgCl₂, 300 µM each of dATP, dCTP and dGTP, 600 µM of dUTP, 100 nM each of forward primer, reverse primer, and probe, 20 U RNase inhibitor, 1.25 units AMPLITAQ GOLD®, and 12.5 U MuLV reverse transcriptase) to 96 well plates containing 25 µl poly(A) mRNA solution. The RT reaction is carried out by incubation for 30 minutes at 48°C. following a 10 minute incubation at 95°C to activate the AMPLITAQ GOLD®, 40 cycles of a two-step PCR protocol are carried out: 95°C for 15 seconds (denaturation) followed by 60°C for 1.5 minutes (annealing/extension).

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STAT3 PCR primers and a probe can be designed using commercial software (e.g. Oligo 5.0).

EXAMPLE 3: Mouse STAT3 Oligonucleotide Sequences

Antisense oligonucleotides were designed to target
5 mouse STAT3. Target sequence data are from the STAT3 cDNA
sequence submitted by Zhong, Z.; Genbank accession number
U06922, provided herein as SEQ ID NO: 82. Oligonucleotides
were synthesized as chimeric oligonucleotides ("gapmers")
20 nucleotides in length, composed of a central "gap"
10 region consisting of ten 2'-deoxynucleotides, which is
flanked on both sides (5' and 3' directions) by
five-nucleotide "wings." The wings are composed of
2'-methoxyethyl (2'-MOE)nucleotides. The internucleoside
(backbone) linkages are phosphorothioate (P=S) throughout
15 the oligonucleotide. All 2'-MOE cytosines were 5-methyl-
cytosines. Oligonucleotide sequences are shown in Table 3.

The B lymphoma cell line, BCL1 was obtained from ATCC
(Rockville, MD). BCL1 cells were cultured in RPMI 1640
medium.

20 BCL1 cells (5×10^6 cells in PBS) were transfected with
oligonucleotides by electroporation, at 200V, 1000 μ F using
a BTX Electro Cell Manipulator 600 (Genetronics, San Diego,
CA). For an initial screen, BCL1 were electroporated with
10 μ M oligonucleotide and RNA collected 24 hours later.
25 Controls without oligonucleotide were subjected to the same
electroporation conditions.

Total cellular RNA was isolated using the RNEASY® kit
(Qiagen, Santa Clarita, CA). RNase protection experiments
were conducted using RIBOQUANT™ kits and template sets
30 according to the manufacturer's instructions (Pharmingen,
San Diego, CA). Northern blotting was performed as
described in Chiang, M-Y. et al. (*J. Biol. Chem.*, 1991,
266, 18162-18171), using a rat cDNA probe prepared by Xho
I/Sal I restriction digest of psvsport-1 plasmid (ATCC,

Rockville, MD). mRNA levels were quantitated using a PhosphorImager (Molecular Dynamics, Sunnyvale, CA).

TABLE 3:
Nucleotide Sequences of Mouse STAT3 Chimeric (deoxy gapped)
Phosphorothioate Oligodeoxynucleotides

5	ISIS NO.	NUCLEOTIDE SEQUENCE ¹ (5' -> 3')	SEQ ID NO:	TARGET GENE NUCLEOTIDE CO-ORDINATES ²	GENE TARGET REGION
	17136	GTTCCACTGAGCCATCCTGC	83	0064-0083	AUG
10	17137	TTCAGGTAGCGTGTGTCCAG	84	0096-0115	coding
	17138	ATGTGACTCTTTGCTGGCTG	85	0205-0224	coding
	17139	CCAAGAGATTATGAAACACC	86	0233-0252	coding
	17140	GCTCCAACATCTGCTGCTTC	87	0485-0504	coding
	17141	GCTCTTCATCAGTCAGTGTC	88	0767-0786	coding
15	17142	ATCTGACACCCTGAGTAGTT	89	1680-1699	coding
	17143	GCCAGACCCAGAAGGAGAAG	90	1742-1761	coding
	17144	CGCTCCTTGCTGATGAAACC	91	1827-1846	coding
	17145	AACTTGGTCTTCAGGTACGG	92	2178-2197	coding
	17146	ATCAATGAATCTAAAGTGCG	93	2253-2272	coding
20	17147	TCAGCACCTTCACCGTTATT	94	2283-2302	coding
	17148	ACTCAAAGTGCCTCCTGCT	95	2309-2328	coding
	17149	GGTTTCAGCTCCTCACATGG	96	2374-2393	STOP
	17150	TAAAAAAAAAATCTGGAAC	97	2485-2504	3'-UTR
	17151	AAGATAGCAGAAGTAGGAAA	98	2506-2525	3'-UTR
25	17152	AAAAAGTGCCAGATTGCC	99	2527-2546	3'-UTR
	17153	ATCACCACACTCACTCATT	100	2557-2645	3'-UTR
	17154	CCTTTGCCTCCCTTCTGCTC	101	2626-2645	3'-UTR
	17155	TGAAAAAGGAGGGCAGGCGG	102	2665-2684	3'-UTR
	17156	CACCAGGAGGCACTTGTCTA	103	2705-2724	3'-UTR
30	17157	AACCTCCTGGGCTTAGTCCT	104	2822-2841	3'-UTR

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23176	AAAAAGTGCGCAGATTGCCC	105	1 base mismatch control
23177	AAAAAGTCCGCTGATTGCCC	106	3 base mismatch control
23178	AAAAACTCCGCTGAATGCCC	107	5 base mismatch control

5 ¹ All 2'-MOE cytosine residues are 5-methyl-cytosines; all linkages are phosphorothioate linkages.

²Co-ordinates from Genbank Accession No. U06922, locus name "MMU06922", SEQ ID NO. 82.

10 Results are shown in Table 4. Oligonucleotides 17138 (SEQ ID NO. 85), 17139 (SEQ ID NO. 86), 17140 (SEQ ID NO. 87), 17143 (SEQ ID NO. 90), 17144 (SEQ ID NO. 91), 17152 (SEQ ID NO. 99), 17153 (SEQ ID NO. 100), 17156 (SEQ ID NO. 103), and 17157 (SEQ ID NO. 104) gave better than 45%
15 inhibition in this assay.

TABLE 4

Inhibition of Mouse STAT3 mRNA expression in BCL1 Cells by Chimeric (deoxy gapped) Phosphorothioate Oligonucleotides

20	ISIS	SEQ ID	GENE	% mRNA	% mRNA
	No:	NO:	TARGET REGION	EXPRESSION	INHIBITION
	control	---	---	100%	0%
	17136	83	AUG	75%	25%
	17137	84	coding	75%	25%
	17138	85	coding	37%	63%
25	17139	86	coding	41%	59%
	17140	87	coding	40%	60%
	17141	88	coding	62%	38%
	17142	89	coding	70%	30%
	17143	90	coding	42%	58%

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	17144	91	coding	55%	45%
	17145	92	coding	89%	11%
	17146	93	coding	91%	9%
	17147	94	coding	70%	30%
5	17148	95	coding	69%	31%
	17149	96	STOP	70%	30%
	17150	97	3'-UTR	95%	5%
	17151	98	3'-UTR	92%	8%
	17152	99	3'-UTR	25%	75%
10	17153	100	3'-UTR	44%	56%
	17154	101	3'-UTR	80%	20%
	17155	102	3'-UTR	78%	22%
	17156	103	3'-UTR	40%	60%
	17157	104	3'-UTR	53%	47%

15

EXAMPLE 4: Dose response of antisense chimeric (deoxy gapped) phosphorothioate oligonucleotide effects on mouse STAT3 protein levels in BCL1 cells

ISIS 17152 (SEQ ID. NO. 99) was chosen for further study. The effect of this oligonucleotide on protein levels was determined by Western blot. ISIS 23177 (SEQ ID NO. 106), a 3 base mismatch, was used as a control. BCL1 cells were grown, treated and processed as described in Example 2.

25 Nuclear extracts from primary B cells and B lymphoma cell lines were prepared as described in Karras, J.G., et al. (*J. Exp. Med.*, 1997, 185, 1035-1042).

Western blotting was performed as described in Karras, J.G. et al. (*J. Immunol.*, 1996, 157, 2299). STAT1 and
30 STAT3 antibodies were obtained from UBI (Lake Placid, NY).

Results are shown in Table 5. ISIS 17152 (SEQ ID NO. 99) was significantly better at reducing STAT3 protein levels than the mismatch control.

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TABLE 5

Dose Response of BCL1 cells to STAT3
Chimeric (deoxy gapped) Phosphorothioate Oligonucleotides

	SEQ ID	ASO Gene		% protein	% protein	
5	ISIS #	NO:	Target	Dose	Expression	Inhibition
	control	---	---	---	100%	---
	17152	99	3'-UTR	10 nM	41.7%	58.3%
	"	"	"	15 nM	42.5%	57.5%
	"	"	"	20 nM	26.5%	73.5%
10	23177	106	control	10 nM	75.1%	24.9%
	"	"	"	15 nM	67.6%	32.4%
	"	"	"	20 nM	62.6%	37.4%

EXAMPLE 5: Inhibition of BCL1 proliferation by STAT3

15 antisense chimeric (deoxy gapped) phosphorothioate oligonucleotide

The effect of ISIS 17152 (SEQ ID NO. 99) on BCL1 proliferation was determined. BCL1 cells contain constitutively active STAT3 which is thought to be responsible for their proliferation. BCL1 cells were grown, treated and processed as described in Example 2.

20 1 X 10⁵ BCL1 cells were incubated in 96-well plates in 200 µL complete RPMI following electroporation. Cultures were pulsed with 1 µCi of [³H]-thymidine for the last 8 hours of culture and cells were harvested and analyzed for thymidine incorporation as described in Francis, D.A. et al. (Int. Immunol., 1995, 7, 151-161) 48 hours after electroporation.

Results are shown in Table 6. ISIS 17152 (SEQ ID NO. 99) was able to reduce BCL1 cell proliferation by approximately 50% whereas the mismatch control had no effect.

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TABLE 6

Inhibition of BCL1 Cell Proliferation with STAT3
Chimeric (deoxy gapped) Phosphorothioate Oligonucleotides

5	SEQ ID	ASO Gene	% Cell		% Cell
	ISIS #	NO: Target	Dose	Proliferation	Inhibition
	control	---	---	100%	---
	17152	99 3'-UTR	10 nM	78.5%	21.5%
	"	"	15 nM	54.4%	45.6%
10	"	"	20 nM	50.2%	49.8%
	23177	106 control	10 nM	117.0%	---
	"	"	15 nM	99.7%	0.3%
	"	"	20 nM	107.0%	---

15 **EXAMPLE 6: Inhibition of BCL1 IgM Secretion by STAT3**
antisense chimeric (deoxy gapped) phosphorothioate
oligonucleotides

The effect of ISIS 17152 (SEQ ID. NO. 99) on IgM
secretion levels was determined. STAT3 has been implicated
20 in regulation of IgM expression (Faris, M., et al.,
Immunology, 1997, 90, 350-357). BCL1 cells were grown,
treated and processed as described in Example 2.

1 X 10⁶ BCL1 cells were incubated in 12-well plates in
2 mL complete RPMI following electroporation. Supernatant
25 was replaced at 24 hour post electroporation with fresh
medium. 48 hours later, supernatants were harvested,
centrifuged to remove cells, and assayed for IgM content
using the OPT-EIA™ ELISA kit (Pharmingen, San Diego, CA)
and capture and detecting antibodies for mouse IgM
30 (Southern Biotechnology, Birmingham, AL).

Results are shown in Table 7. ISIS 17152 (SEQ ID NO.
99) was significantly better at reducing IgM secretion than
the mismatch control.

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TABLE 7

Inhibition of BCL1 IgM secretion by STAT3
Chimeric (deoxy gapped) Phosphorothioate Oligonucleotides

5	SEQ ID	ASO Gene	% IgM		% IgM	
	ISIS #	NO:	Target	Dose	Expression	Inhibition
10	control	---	---	---	100%	---
	17152	99	3'-UTR	5 nM	34.2%	65.8%
	"	"	"	15 nM	23.1%	76.9%
	23177	106	control	5 nM	110.0%	---
	"	"	"	15 nM	80.8%	19.2%

EXAMPLE 7: Induction of Chemokines in BCL1 cells following
Treatment with STAT3 antisense chimeric (deoxy gapped)

15 phosphorothioate oligonucleotide

The effect of ISIS 17152 (SEQ ID. NO. 99) on chemokine levels was determined. BCL1 cells were grown, treated and processed as described in Example 2. Chemokine gene expression was induced in BCL1 cells by addition of 10 μ M of a CpG-containing oligonucleotide to the media 16 hours following antisense oligonucleotide electroporation. CpG-containing oligonucleotides are immune-stimulatory (Krieg, A.M., et al., *Nature*, 1995, 374, 546-549). The levels of chemokines were measured eight hours later using RNase protection assay as described in Example 2 with a mouse chemokine template set, Mck-5 (Pharmingen, San Diego, CA).

Results are shown in Table 8. ISIS 17152 (SEQ ID. NO. 99) was able to induce the expression of the chemokines, RANTES, MIP-1 α and MIP-1 β whereas the mismatch control had minimal effect.

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TABLE 8

Induction of Chemokines in BCL1 Cells Following Treatment
with STAT3 Chimeric (deoxy gapped) Phosphorothioate
Oligonucleotides

5	SEQ	ASO Gene		%	% MIPl _a	% MIPl _b
	ISIS #	ID	Target	Dose	mRNA	mRNA
	NO:			mRNA		
	control	---	---	---	100%	100%
	17152	99	3'-UTR	5 nM	236%	201%
	"	"	"	10 nM	266%	258%
10	"	"	"	20 nM	257%	254%
	23178	107	control	5 nM	96%	123%
	"	"	"	10 nM	70.2%	116%
	"	"	"	20 nM	56%	106%

15 **EXAMPLE 8: Effect of STAT3 Antisense Oligonucleotides in a Murine Model for Rheumatoid Arthritis**

Collagen-induced arthritis (CIA) is used as a murine model for arthritis (Mussener, A., et al., *Clin. Exp. Immunol.*, 1997, 107, 485-493). Female DBA/1LacJ mice
20 (Jackson Laboratories, Bar Harbor, ME) between the ages of 6 and 8 weeks are used to assess the activity of STAT3 antisense oligonucleotides.

On day 0, the mice are immunized at the base of the tail with 100 µg of bovine type II collagen which is
25 emulsified in Complete Freund's Adjuvant (CFA). On day 7, a second booster dose of collagen is administered by the same route. On day 14, the mice are injected subcutaneously with 100 µg of LPS. Oligonucleotide is administered intraperitoneally daily (10 mg/kg bolus)
30 starting on day -3 and continuing for the duration of the study.

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Weights are recorded weekly. Mice are inspected daily for the onset of CIA. Paw widths are rear ankle widths of affected and unaffected joints and are measured three times a week using a constant tension caliper. Limbs are

5 clinically evaluated and graded on a scale from 0-4 (with 4 being the highest).

Example 9: Effect of STAT3 antisense oligonucleotides on growth of human MDA-MB231 tumors in nude mice

MDA-MB231 human breast carcinoma cells are obtained

10 from the American Type Culture Collection (Bethesda, MD). Serially transplanted MDA-MB231 tumors are established subcutaneously in nude mice. Beginning two weeks later, STAT3 antisense oligonucleotides, in saline, are administered intravenously daily for 14 days at dosages of

15 60 mg/kg and 6 mg/kg. Control oligonucleotides are also administered at these doses, and a saline control is also given. Tumor growth rates are monitored for the two-week period of oligonucleotide administration. Activity of the STAT3 antisense oligonucleotides is measured by a reduction

20 in tumor growth. A lower-dose study can also be conducted using the same oligonucleotides at 6 mg/kg and 0.6 mg/kg.

Example 10: Effect of STAT3 antisense oligonucleotides on U-87 human glioblastoma cells following subcutaneous xenografts into nude mice:

25 The U-87 human glioblastoma cell line is obtained from the ATCC (Rockville MD) and maintained in Iscove's DMEM medium supplemented with heat-inactivated 10% fetal calf serum. Nude mice are injected subcutaneously with 2×10^7 cells. Mice are injected intraperitoneally with STAT3

30 antisense oligonucleotides at dosages of either 2 mg/kg or 20 mg/kg for 21 consecutive days beginning 7 days after xenografts are implanted. Tumor volumes are measured on days 14, 21, 24, 31 and 35. Activity is measured by

-62-

reduced tumor volume compared to saline or sense oligonucleotide control.

Example 11: Effect of STAT3 antisense oligonucleotides on intracerebral U-87 glioblastoma xenografts into nude mice

5 U-87 cells are implanted in the brains of nude mice. Mice are treated via continuous intraperitoneal administration of STAT3 antisense oligonucleotides at 20 mg/kg, control sense oligonucleotide (20 mg/kg) or saline beginning on day 7 after xenograft implantation. Activity
10 of the STAT3 antisense oligonucleotides is measured by an increase in survival time compared to controls.

Example 12: Additional antisense oligonucleotides targeted to human STAT3

An additional set of oligonucleotides targeted to SEQ ID
15 NO: 1 was designed and synthesized as chimeric oligonucleotides ("gapmers") 20 nucleotides in length, composed of a central "gap" region consisting of ten 2'-deoxynucleotides, which is flanked on both sides (5' and 3' directions) by five-nucleotide "wings." The wings are
20 composed of 2'-methoxyethyl (2'-MOE)nucleotides (shown in bold). The internucleoside (backbone) linkages are phosphorothioate (P=S) throughout the oligonucleotide. All 2'-MOE cytosines and 2'-deoxy cytosines were 5-methyl-cytosines. These oligonucleotide sequences are shown in
25 Table 9.

TABLE 9:

Nucleotide Sequences of Additional Chimeric (deoxy gapped) Phosphorothioate Oligonucleotides targeted to Human STAT3

30	ISIS NO.	NUCLEOTIDE SEQUENCE ¹ (5' -> 3')	TARGET GENE NUCLEOTIDE CO- ORDINATES ²	GENE TARGET REGION	SEQ ID NO:
	113169	ATGTGATTCTTTGCTGGCCG	357	5' UTR	108
	113170	AGCTGATTCCATTGGGCCAT	221	AUG	109
	113171	CCAGGAGATTATGAAACACC	385	Coding	110

30	ISIS NO.	NUCLEOTIDE SEQUENCE ¹ (5' -> 3')	TARGET GENE NUCLEOTIDE CO- ORDINATES ²	GENE TARGET REGION	SEQ ID NO:
	113172	ACCGTGTGTCAAGCTGCTGT	241	Coding	111
	113173	CCATTGGGAAGCTGTCCTG	286	Coding	112
	113174	TGTGATTCTTTGCTGGCCGC	356	Coding	113
	113175	GCGGCTATACTGCTGGTCAA	411	Coding	114
5	113176	GCTCCAGCATCTGCTGCTTC	637	Coding	115
	113177	GATTCTTCCCACAGGCACCG	539	Coding	116
	113178	TGATTCTTCCCACAGGCACC	540	Coding	117
	113179	ATCCTGAAGGTGCTGCTCCA	651	Coding	118
	113180	CGGACATCCTGAAGGTGCTG	656	Coding	119
10	113181	CCCGCCAGCTCACTCACGAT	869	Coding	120
	113182	AGTCAGCCAGCTCCTCGTCC	928	Coding	121
	113183	CCAGTCAGCCAGCTCCTCGT	930	Coding	122
	113184	CGCCTCTTCCAGTCAGCCAG	938	Coding	123
	113195	GGCCGGTGCTGTACAATGGG	1109	Coding	124
15	113186	ATCCTCTCCTCCAGCATCGG	1127	Coding	125
	113187	CCGCTCCACCACAAAGGCAC	1176	Coding	126
	113188	CGTCCCCAGAGTCTTTGTCA	1324	Coding	127
	113189	TTGTGTTTGTGCCAGAATG	1375	Coding	128
	113190	GCTCGGCCCCCATTCACACA	1472	Coding	129
20	113191	AGGCATTTGGCATCTGACAG	1621	Coding	130
	113192	CTTGGGATTGTTGGTCAGCA	1665	Coding	131
	113193	CTCGGCCACTTGGTCCCAGG	1719	Coding	132
	113194	CCCCGCTTGGTGGTGGACGA	1757	Coding	133
	113195	CCCCGCTTGGTGGTGGACG	1758	Coding	134
25	113196	GGAGAAGCCCTTGCCAGCCA	1881	Coding	135
	113197	TTCATTCCAAAGGGCCAAGA	1947	Coding	136
	113198	CCCGCTCCTTGCTGATGAAA	1881	Coding	137
	113199	GTGCTCAAGATGGCCCGCTC	2000	Coding	138
	113200	CCCAAGTGAAAGTGACGCCT	2071	Coding	139
30	113201	ACCCAAGTGAAAGTGACGCC	2072	Coding	140
	113202	CCGAATGCCTCCTCCTTGGG	2252	Coding	141
	113203	GCCGACAATACTTCCCGAAT	2266	Coding	142
	113204	GATGCTCCTGGCTCTCTGGC	2284	Coding	143
	113205	TCAATGAATCTAAAGCGCGG	2404	Coding	144

30	ISIS NO.	NUCLEOTIDE SEQUENCE ¹ (5' -> 3')	TARGET GENE NUCLEOTIDE CO- ORDINATES ²	GENE TARGET REGION	SEQ ID NO:
	113206	GACTCAAAGTGGCCCTCCTGC	2462	Coding	145
	113207	ATCACCCACATTCACTCATT	2710	3' UTR	146
	113208	AAAAGTGCCCGAGATTGC	2682	3' UTR	147
	113209	AAAAGTGCCCGAGATTGCTCA	2679	3' UTR	148
5	113210	TAAAAGTGCCCGAGATTGCTC	2680	3' UTR	149
	113211	AAGCAGATCACCCACATTCA	2716	3' UTR	150

These oligonucleotides were screened by Northern blot analysis in U266 cells at an oligonucleotide concentration of 2.5 μ M. U266 human myeloma cell lines (originally obtained from American Type Culture Collection) were maintained in RPMI 1640 medium supplemented with 10% fetal calf serum. Cells (15×10^6 cells in PBS) were transfected with oligonucleotides at 200V with a single 6-millisecond pulse using a BTX Electro Square Porator T820 (Genetronics, San Diego CA). The cells were incubated for 24 hours before RNA extraction.

Total cellular RNA was isolated using the Rneasy kit (Qiagen, Santa Clarita, CA). Northern blotting was performed on 15 μ g of RNA using a cDNA probe prepared from MB-MDA 468 RNA by standard RT-PCR followed by a nested primer reaction. Signals were quantitated using a Molecular Dynamics Phosphorimager.

Results for selected compounds (expressed as percent of control mRNA expression and percent inhibition of mRNA expression) are shown in Table 10.

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TABLE 10

Inhibition of Human STAT3 mRNA expression in U266 Cells by
Chimeric (deoxy gapped) Phosphorothioate Oligonucleotides

5	ISIS No:	SEQ ID NO:	GENE TARGET REGION	% mRNA EXPRESSION	% mRNA INHIBITION
	None	--	--	100	--
	17148	95 ✓	Coding	95.1	4.9
	17152	99	3' UTR	82.5	17.5
	113170	109	AUG	89.6	10.4
10	113171	112	Coding	110.2	--
	113172	111	Coding	96.1	3.9
	113173	112	Coding	119	--
	113175	114	Coding	75.8	24.2
	113176	115 ✓	Coding	72.3	27.7
15	113178	117	Coding	143.9	--
	113181	120	Coding	105.4	--
	113184	123	Coding	104.3	--
	113187	126 ✓	Coding	55.9	44.1
	113189	128	Coding	163.9	--
20	113199	139	Coding	64.4	35.6
	113207	146	3' UTR	123.6	--
	113209	148	3' UTR	71.4	28.6
	113210	149 ✓	3' UTR	72.2	27.8
25	113211	150	3' UTR	116.5	--

Dose-response experiments were conducted for ISIS 113176, 129987, 113187, 129991, 113209, 129995, 113210 and 129999 as well as ISIS 17148 and the mouse STAT3 oligo ISIS 114054. Results are shown in Table 11.

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Table 11

Percent inhibition of human STAT3 mRNA expression with
antisense oligonucleotides- dose response

5	ISIS #	SEQ ID NO:	Percent inhibition of STAT3 expression		
			Oligo concentration		
			2.5 μ M	5 μ M	10 μ M
	17148	95	8	54	60
	114054		4	17	15
	113176	115	33	67	79
	129987		5	5	29
10	113187	126	44	56	75
	129991		21	22	26
	113209	148	43	54	73
	129995		5	32	25
	113210	149	36	50	76
15	129999		31	8	--

ISIS 17148, 113176, 113187, 113209 and 113210 were shown to
reduce STAT3 expression by over 50% at one or more
oligonucleotide concentrations. These compounds are
20 therefore preferred.

**Example 13: Antisense inhibition of STAT3 causes apoptotic
cell death in mouse melanoma cells**

Mouse B16 melanoma cells were grown in RPMI 1640 (Life
Technologies, Inc., Grand Island, NY) medium supplemented
25 with 10% fetal bovine serum, 2 mM L-glutamine, 1 mM sodium
pyruvate, 1% MEM nonessential amino acids and 100 IU/ml
penicillin/streptomycin.

Cells were treated with ISIS 17152, targeted to mouse
STAT3, or the 3-base mismatch control, ISIS 28084
30 (AAAAAGAGGCCTGATTGCCC; SEQ ID NO: 151). Cells were

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transfected with oligonucleotide using LipofectAMINE PLUS™ reagent (GibcoBRL). Oligonucleotide was pre-complexed with LipofectAMINE PLUS™ by adding the oligonucleotide to 100 μ l serum-free RPMI 1640 medium, then 6 μ l LipofectAMINE PLUS™ reagent was added, the sample was mixed well and incubated for 15 minutes at room temperature. An additional 4 μ l of LipofectAMINE PLUS™ reagent was diluted to 100 μ l in serum-free RPMI. This diluted LipofectAMINE PLUS™ was mixed with the pre-complexed oligonucleotide/LipofectAMINE PLUS™ mixture and incubated for 15 minutes at room temperature. 800 μ l of serum-free RPMI 1640 was added, and the resulting oligonucleotide-LipofectAMINE PLUS™-medium mixture (approximately 1 ml) was added to cells in a 6-well plate. After 3 hours incubation, 1 ml of RPMI 1640 supplemented with 20% fetal bovine serum was added. Oligonucleotide concentrations were 200 nM or 300 nM.

24 hours after transfection, cells were counted to determine the effect of antisense treatment on cell death. Cells were harvested at 24 hours post transfection for western blot analysis and at 48 hours post-transfection for Annexin-V staining for apoptosis.

Effects of oligonucleotide on cell number are shown in Table 12.

Table 12

Effect of antisense inhibition of STAT3 on cell number

Expt	200 nM		300 nM	
	ISIS 28084 (3 mismatch)	ISIS 17152	ISIS 28084 (3 mismatch)	ISIS 17152
1	10.2×10^5	3.8×10^5		
2	5.0×10^5	6.8×10^5	9.1×10^5	3.5×10^5
3	3.5×10^5	1.8×10^5	3.3×10^5	2.2×10^5

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Thus treatment with STAT3 antisense oligonucleotide increased cell death (decreased cell number).

Apoptosis in B16 cells was measured by staining with Annexin V-PE (Clontech) and flow cytometry analysis 48 hours after antisense treatment. Positive staining for Annexin-V indicates apoptosis is occurring. Mock-transfected cells and control oligonucleotide-treated cell cultures had 11.37% and 10.15% of cells staining positive for Annexin-V. In contrast, ISIS 17152-treated cells were 29.84% positive for Annexin-V, indicating a nearly threefold increase in apoptotic cells. It should be noted that in general, the percent of apoptosis in B16 cells is likely to have been underestimated since detached dead cells are washed off in processing.

Western blot analysis was done on cells 24 hours after antisense treatment, using an anti-STAT3 antibody (K15, Santa Cruz Biotechnology, Santa Cruz, CA). ISIS 17152 at 200nM or 300 nM significantly reduced STAT3 protein production in B16 cells.

Example 14: Effect of STAT3 antisense oligonucleotides on melanoma tumors

Six-week-old female C57BL mice were purchased from the National Cancer Center (Frederick MD) and maintained under approved conditions. Mice were shaved in the left flank area and injected subcutaneously with 2×10^5 B16 melanoma cells in 100 μ l of PBS. After 7-10 days, B16 tumors with a diameter of 3-6 mm were established. Tumor volume was calculated according to the formula $V = 0.52 \times a^2 \times b$ (a, smallest superficial diameter; b, largest superficial diameter).

Beginning two weeks later, STAT3 antisense oligonucleotides, in saline, are administered intravenously daily for 14 days at dosages of 60 mg/kg and 6 mg/kg.

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Control oligonucleotides are also administered at these doses, and a saline control is also given. Tumor growth rates are monitored for the two-week period of oligonucleotide administration. Activity of the STAT3
5 antisense oligonucleotides is measured by a reduction in tumor growth. A lower-dose study can also be conducted using the same oligonucleotides at 6 mg/kg and 0.6 mg/kg.

What is claimed is:

1. An antisense compound 8 to 30 nucleobases in length targeted to a nucleic acid molecule encoding STAT3, wherein said antisense compound inhibits the expression of said STAT3.
- 5 2. The antisense compound of claim 1 which is an antisense oligonucleotide.
3. The antisense compound of claim 2 wherein the antisense oligonucleotide comprises at least one modified internucleoside linkage.
- 10 4. The antisense compound of claim 3 wherein the modified internucleoside linkage is a phosphorothioate linkage.
5. The antisense compound of claim 2 wherein the antisense oligonucleotide comprises at least one modified sugar moiety.
6. The antisense compound of claim 5 wherein the modified
15 sugar moiety is a 2'-O-methoxyethyl moiety.
7. The antisense compound of claim 2 wherein the antisense oligonucleotide comprises at least one modified nucleobase.
8. The antisense compound of claim 7 wherein modified nucleobase is a 5-methyl cytosine.
- 20 9. The antisense compound of claim 1 wherein the antisense oligonucleotide is a chimeric oligonucleotide.
10. A pharmaceutical composition comprising the antisense compound of claim 1 and a pharmaceutically acceptable carrier or diluent.
- 25 11. The pharmaceutical composition of claim 10 further comprising a colloidal dispersion system.
12. The pharmaceutical composition of claim 10 wherein the antisense compound is an antisense oligonucleotide.

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13. A method of inhibiting the expression of STAT3 in cells or tissues comprising contacting said cells or tissue with the antisense compound of claim 1 so that expression of STAT3 is inhibited.
- 5 14. A method of treating an animal having a disease or condition associated with STAT3 comprising administering to said animal a therapeutically or prophylactically effective amount of the antisense compound of claim 1 so that expression of STAT3 is inhibited.
- 10 15. The method of claim 14 wherein the disease or condition is an inflammatory or autoimmune disease.
16. The method of claim 15 wherein said inflammatory or autoimmune disease or condition is rheumatoid arthritis.
17. The method of claim 14 wherein said disease or condition
15 is cancer.
18. The method of claim 17 wherein said cancer is a cancer of the breast, prostate, brain, head and/or neck or a leukemia, myeloma, melanoma or lymphoma.
19. A method of inducing apoptosis in a cell comprising
20 contacting a cell with with the antisense oligonucleotide of claim 1, so that apoptosis is induced.
20. A method of treating a human having a disease or condition characterized by a reduction in apoptosis comprising administering to a human a prophylactically or therapeutically
25 effective amount of the antisense oligonucleotide of claim 1.

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INTERNATIONAL SEARCH REPORT

International application No.
PCT/US00/09054

A. CLASSIFICATION OF SUBJECT MATTER

IPC(7) : C07H 21/04; A61K 48/00; C12N 15/85

US CL : 514/44; 435/6, 91.1, 325, 366; 536/23.1, 24.5

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

U.S. : 514/44; 435/6, 91.1, 325, 366; 536/23.1, 24.5

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched
NONEElectronic data base consulted during the international search (name of data base and, where practicable, search terms used)
WEST, MEDLINE, CAPLUS, EMBASE, SCISEARCH

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	BRANCH. A good antisense molecule is hard to find. TIBS. February 1998, Vol. 23, pages 45-50, see entire document.	1-20
A	GEWIRTZ et al. Facilitating oligonucleotide delivery: Helping antisense deliver on its promise. Proc. Natl. Acad. Sci. April 1996, Vol. 93, pages 3161-3163, see entire document.	1-20
A	AGRAWAL. Antisense oligonucleotides: towards clinical trials. TIBTECH. October 1996, Vol. 14, pages 376-387, see entire document.	1-20

☐ Further documents are listed in the continuation of Box C. ☐ See patent family annex.

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